



Ohio EPA HAB Response and Lessons Learned

Nutrients & Harmful Algae Webinar
May 25, 2017

Heather Raymond
Ohio EPA
Division of Drinking and Ground Waters



Outline

- Ohio HAB Response
- Lessons Learned:
 - HABs are Not Always Visually Apparent
 - HABs Occur Year Round
 - Widespread Occurrence & Can Increase Rapidly
 - Severity Can Vary Year to Year
 - Saxitoxins are an Emerging Issue
 - Multi-parameter Datasondes are Useful
 - Analytical Methods Matter
- Ohio HAB Initiatives

Summary of Ohio HAB Response

2010: Ohio EPA began incident-response sampling for cyanotoxins

- Posted recreational advisory signs
- Finished water microcystins detection

2011: Ohio EPA/ODNR/ODH created Ohio HAB Response Strategy

- Established Cyanotoxin Thresholds, Sampling Approach, Advisory Language, Public Notice Templates
- Sample for Microcystins and Utilize Phytoplankton ID to Target Sampling for Saxtoxins, Cylindrospermopsin, and Anatoxin-a

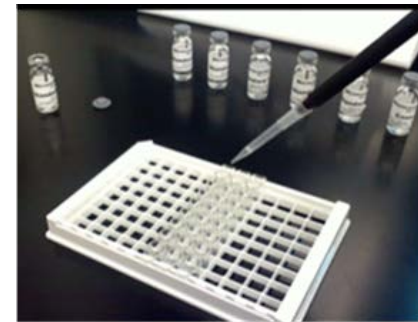
2013 & 2014: Drinking Water Advisories due to Microcystins

2015: USEPA Established Health Advisories for Microcystins and Cylindrospermopsin & Ohio Senate Bill 1 passed

- Finished Water Microcystins Detections at 5 Public Water Systems
- Finished Water Saxitoxins Detections at ? PWSs
- No Drinking Water Advisories!

2016: HAB Monitoring and Reporting Rules Effective June, 2016

- Updated response strategies, new treatment optimization and general plan guidance for PWSs



Ohio HAB Rules Overview

HAB Rules: epa.ohio.gov/ddagw/rules.aspx

- PWS requirements - new rules in OAC Chapter 3745-90
 - Microcystins action levels in drinking water
 - Monitoring requirements: Microcystins and Cyanobacteria Screening
 - Treatment technique requirements
 - Public notification and Consumer Confidence Report (CCR) requirements
 - Recordkeeping requirements
- Laboratory Certification requirements –
New OAC rule 3745-90-04 and amended rules in Chapter 3745-89
 - Laboratory certification
 - Analytical techniques
 - Reporting deadlines



epa.ohio.gov/Portals/28/documents/labcert/TotalMicrocystins.pdf

Ohio Monitoring Requirements

- Total Microcystins

May – October

- Weekly raw and finished water
 - Eligible for monitoring reductions starting May 2017
- Raw water detections >5 ug/L and any finished water detections trigger additional sampling

November – April

- Raw water only every other week
- Detections trigger additional monitoring

- Cyanobacteria Screening (qPCR)

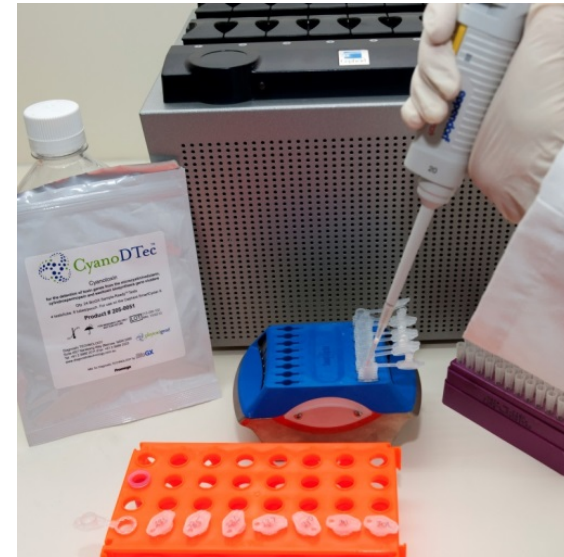
All year

- Biweekly raw water
- Triggers follow up sampling by Ohio EPA for other cyanotoxins



Cyanobacteria Screening: Molecular Methods (Multiplex qPCR)

- Quantitative polymerase chain reaction (qPCR) – identifies and quantifies the presence of genes unique to:
 - Cyanobacteria (16S rDNA)
 - Microcystin and Nodularin production (mcyE gene)
 - Cylindrospermopsin production (cyrA gene)
 - Saxitoxin production (sxtA gene)
 - Test completed within 2-3 hours
 - Scalable
 - Cost-effective
 - Utilizes certified reference material
 - Specific
- Ohio EPA method and certification in 2017
- Ohio EPA uses the data to trigger saxitoxins and cylindrospermopsin sampling and in 2017 will be used as trigger for microcystins monitoring.
- www.phytoxigene.com/products/



Rule Implementation & Monitoring Results

- **100% compliance with rules!**
- Microcystins detected in raw water at 45 PWSs (38%) and mcxE gene detected at 57 PWSs (48%)
- No finished water microcystins detections
- Saxitoxin genes (sxtA) detected in 33 PWSs (27%)
- Saxitoxins detected in raw water at 15 PWSs (12%)
- 6 finished water saxitoxins detections (none above threshold)
- Cylindrospermopsin genes detected at 1 PWS, no cylindrospermopsin detections.

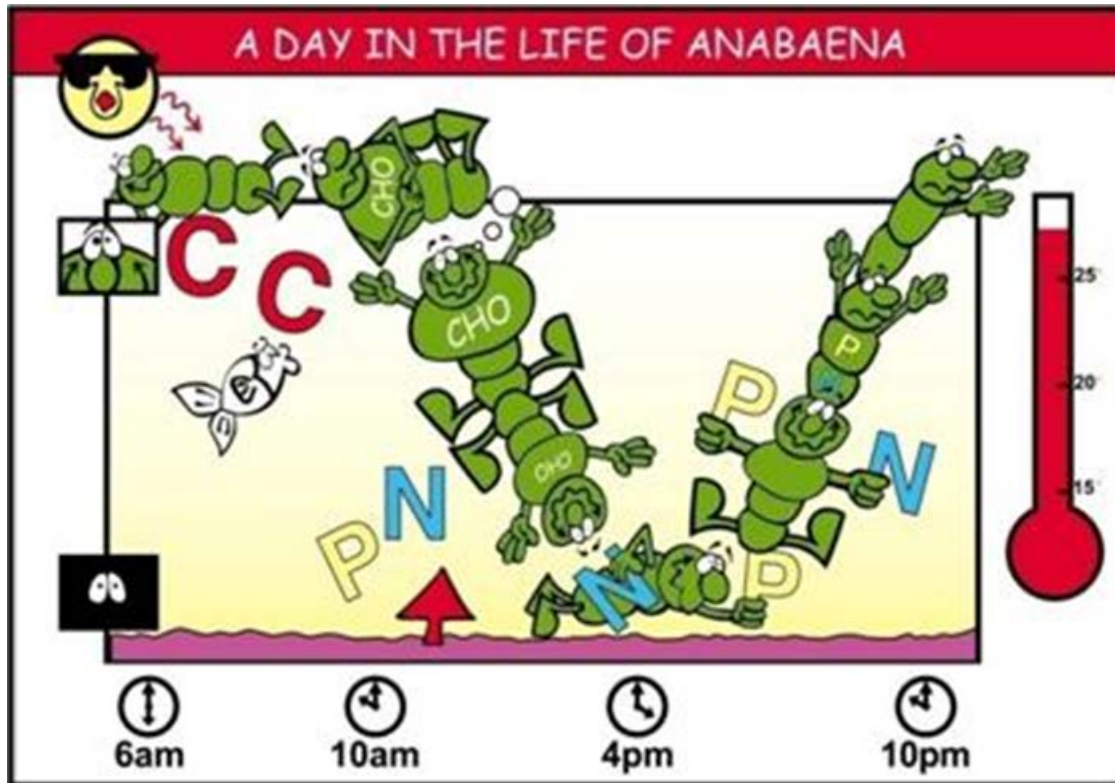


qPCR as a Screening Tool

- 100% of microcystins >1.6 µg/L had paired mcyE gene detections
- 100% of microcystins >5 µg/L had mcyE detections > 5gc/µL
- 90% of microcystins >1.6 ug/L had mcyE detections >5 gc/µL
- Less than 2% of samples had microcystins detections without corresponding mcyE detections
- Less than 1% percent of samples had saxitoxins detections without corresponding gene detections
- Only one cyrA detection, no cylindrospermopsin detections
- Overall, qPCR is effective screening tool for microcystins and saxitoxins and performs better than phytoplankton enumeration.

Lessons Learned – HABs are Not Always Visually Apparent

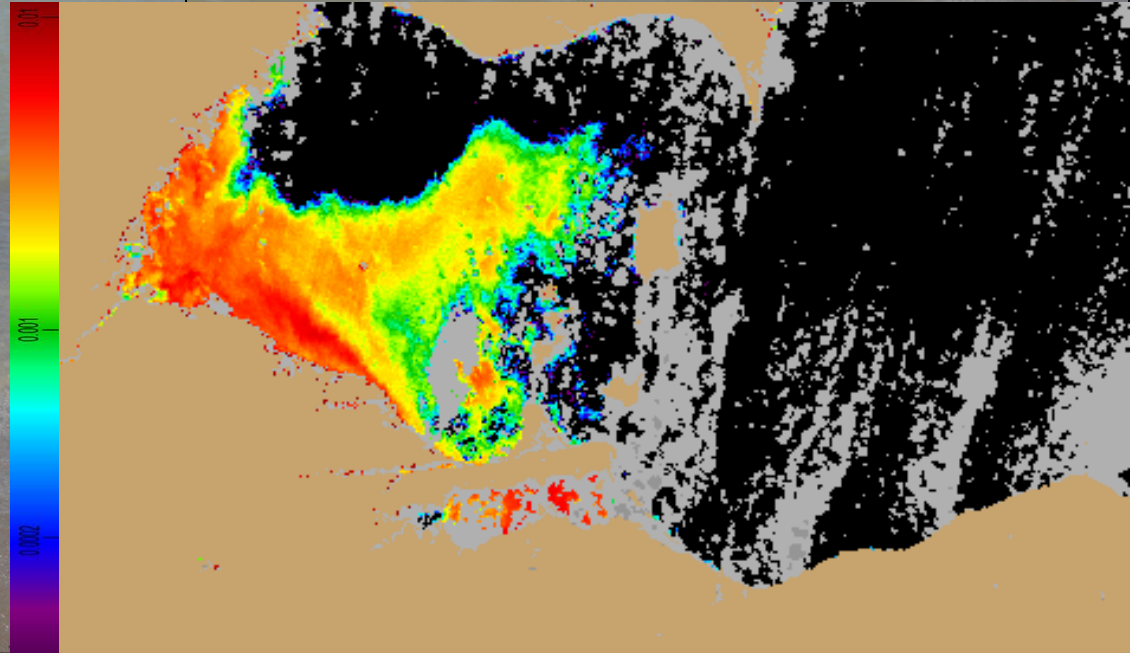
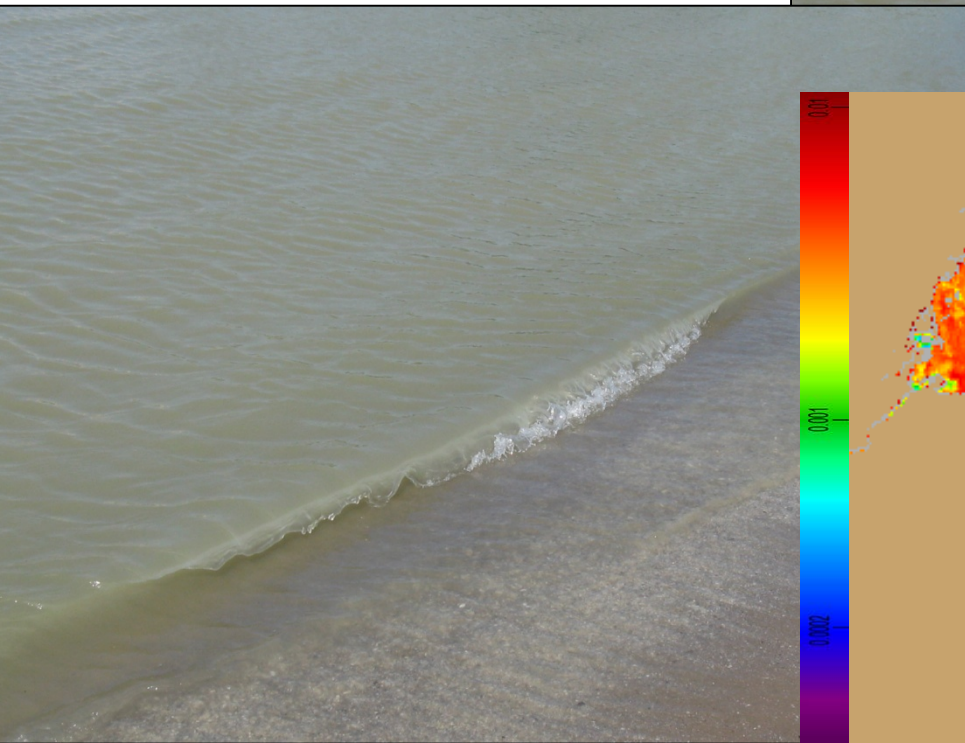
- Many cyanobacteria do not produce surface scums
- Some cyanobacteria occur at depth in the water column or attach to a substrate (benthic)
- Some cyanobacteria move up and down in the water column



Complicates Monitoring

Lake Erie Beach
Microcystis bloom
August 31, 2011

Microcystins >100 ug/L

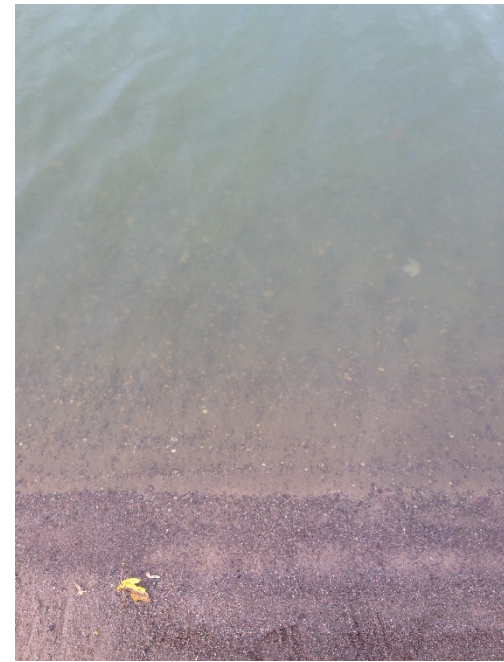




Celina Intake on Grand Lake Saint Mary,
September 2015, *Planktothrix* bloom

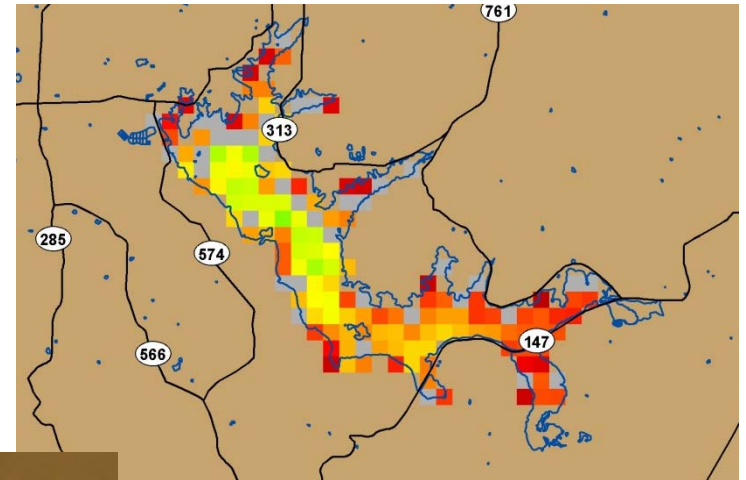
Microcystins 185 ug/L



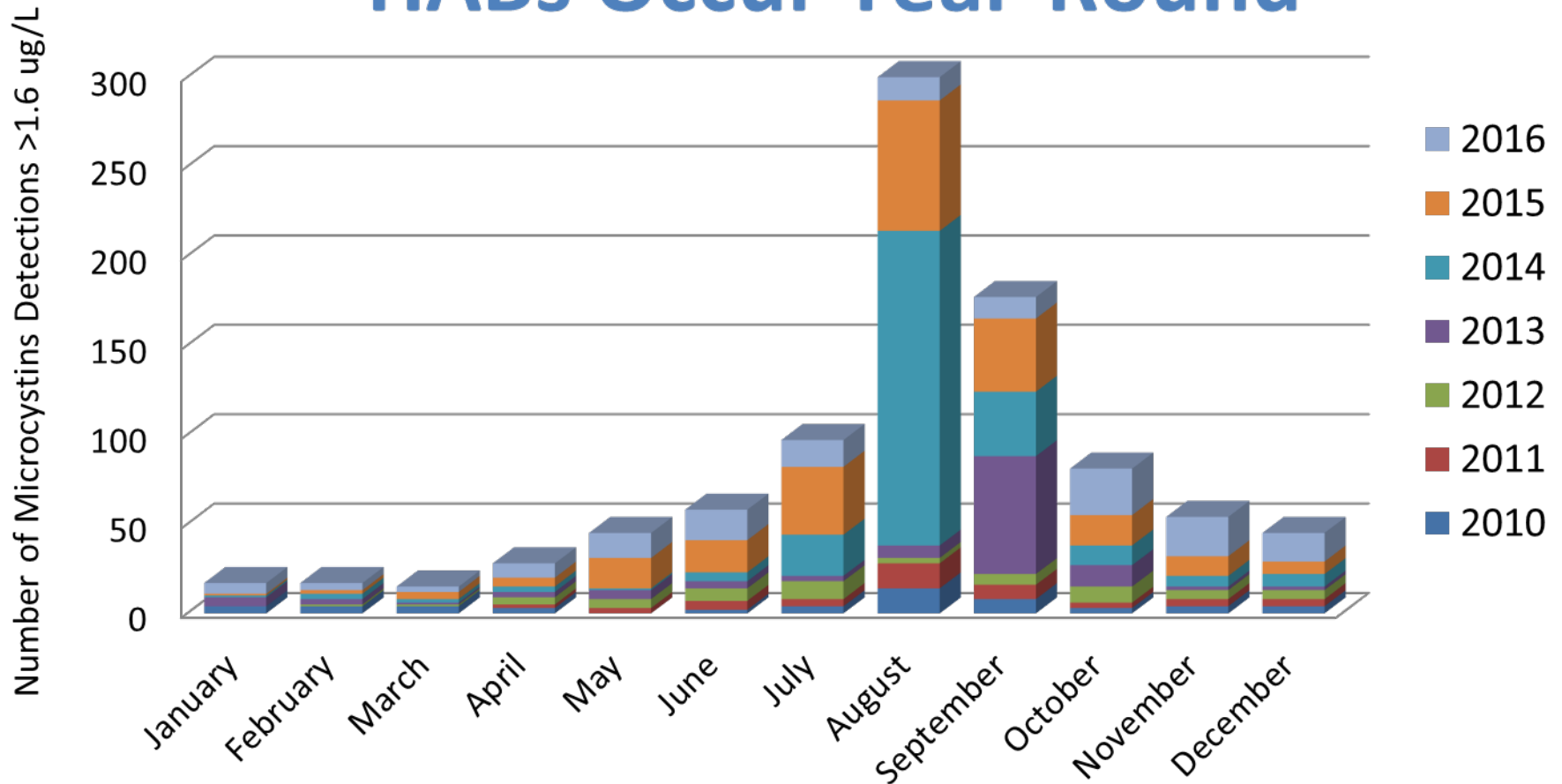


Chippewa Lake, November 2016
Aphanizomenon and *Planktothrix* bloom

Average Microcystins at Beaches 48-58 ug/L



Lessons Learned – HABs Occur Year-Round



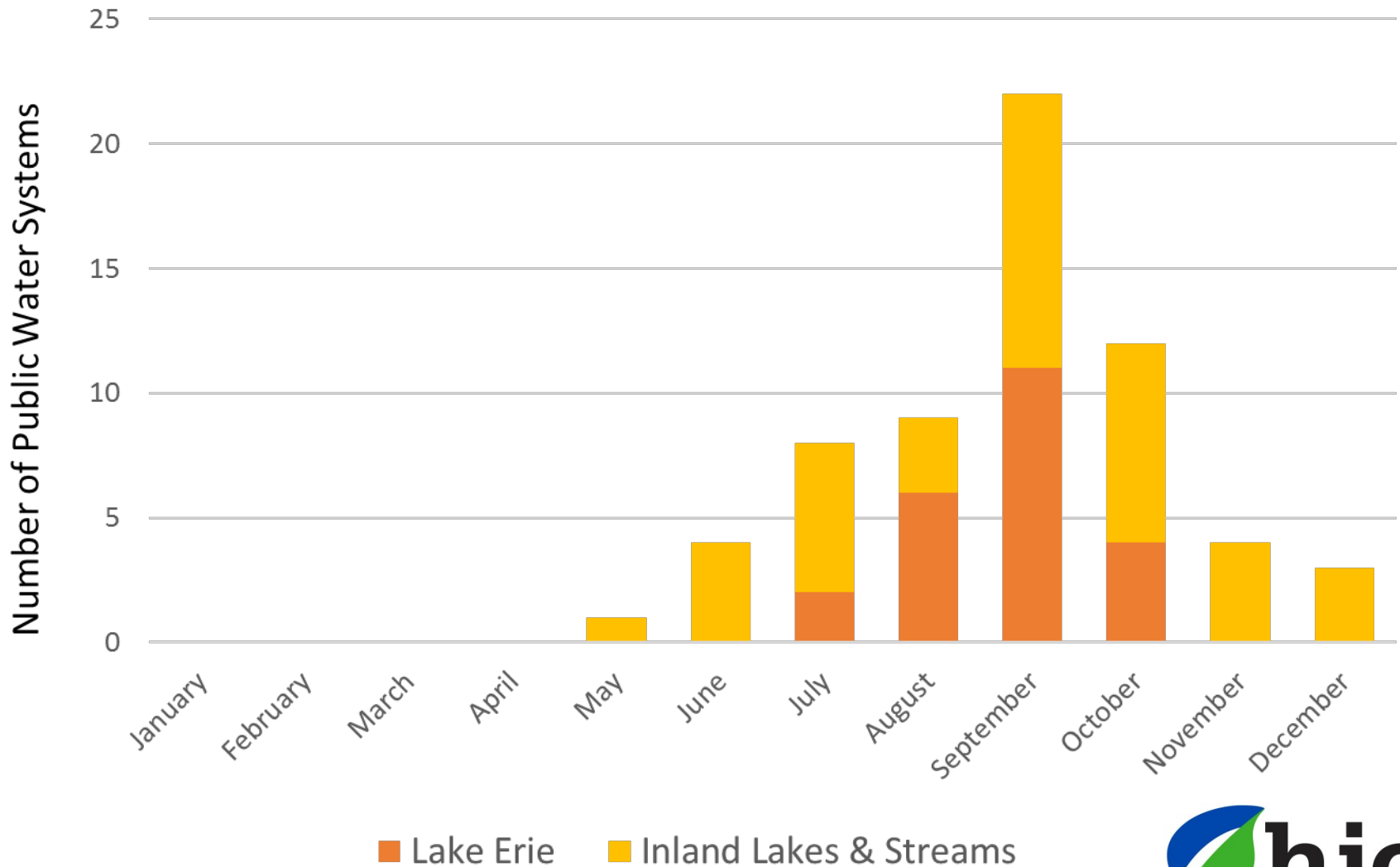
Response Sampling (2010-2015):

- 745 samples >1.6 ug/L microcystins, out of 3583 total samples (21%).
- 44% of samples were > 0.30 ug/L microcystins.

Routine Monitoring (2016):

- 164 samples >1.6 ug/L microcystins, out of 3512 total samples (5%)
- 11% of samples were > 0.30 ug/L microcystins.

Maximum Microcystins Detection By Month



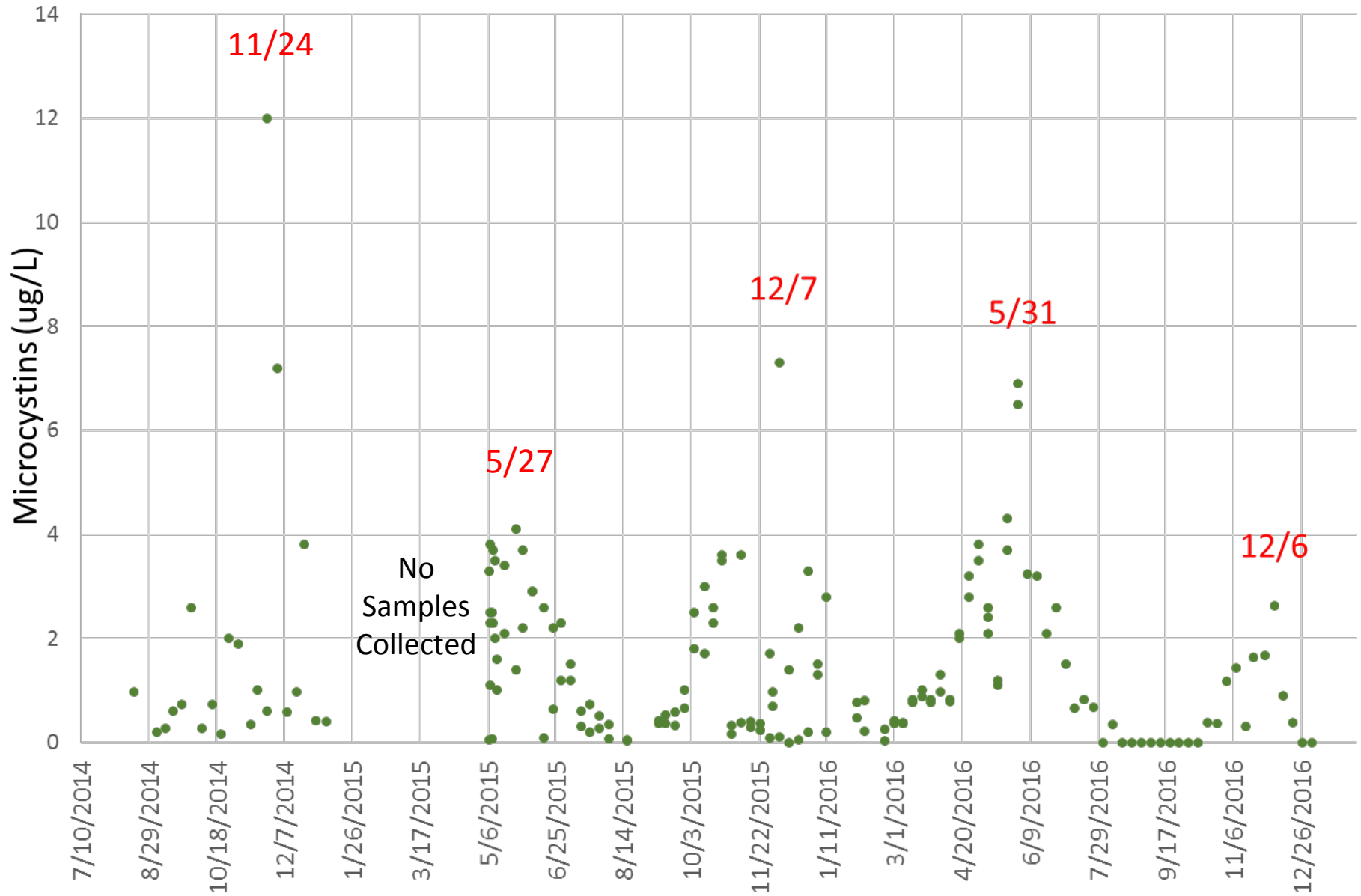
Based on Microcystins Detections at 62 Public Water Systems
(2010 - 2016)



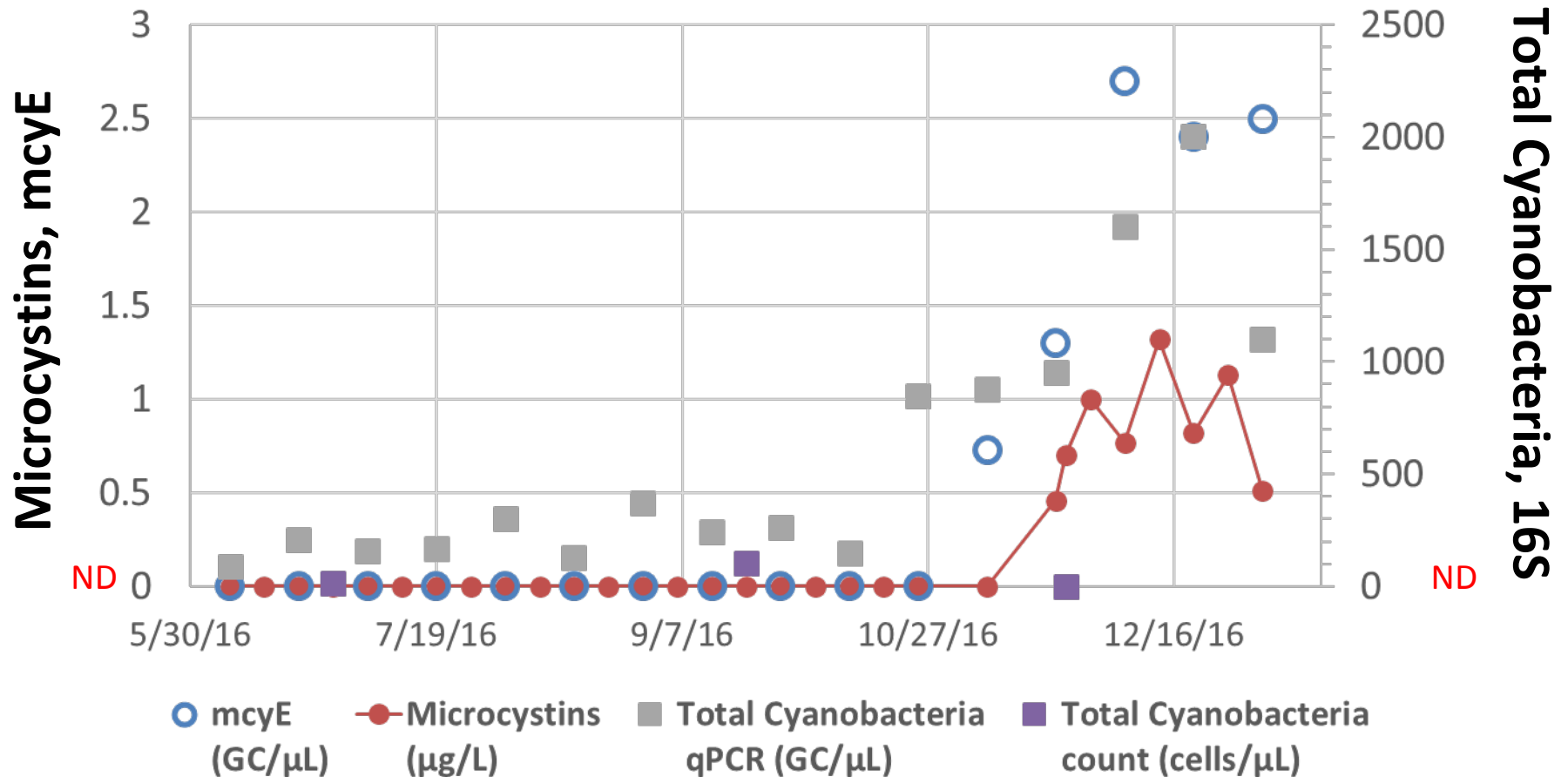
William's Reservoir November, 2012

**Microcystin
Concentration: 1400 ug/L**

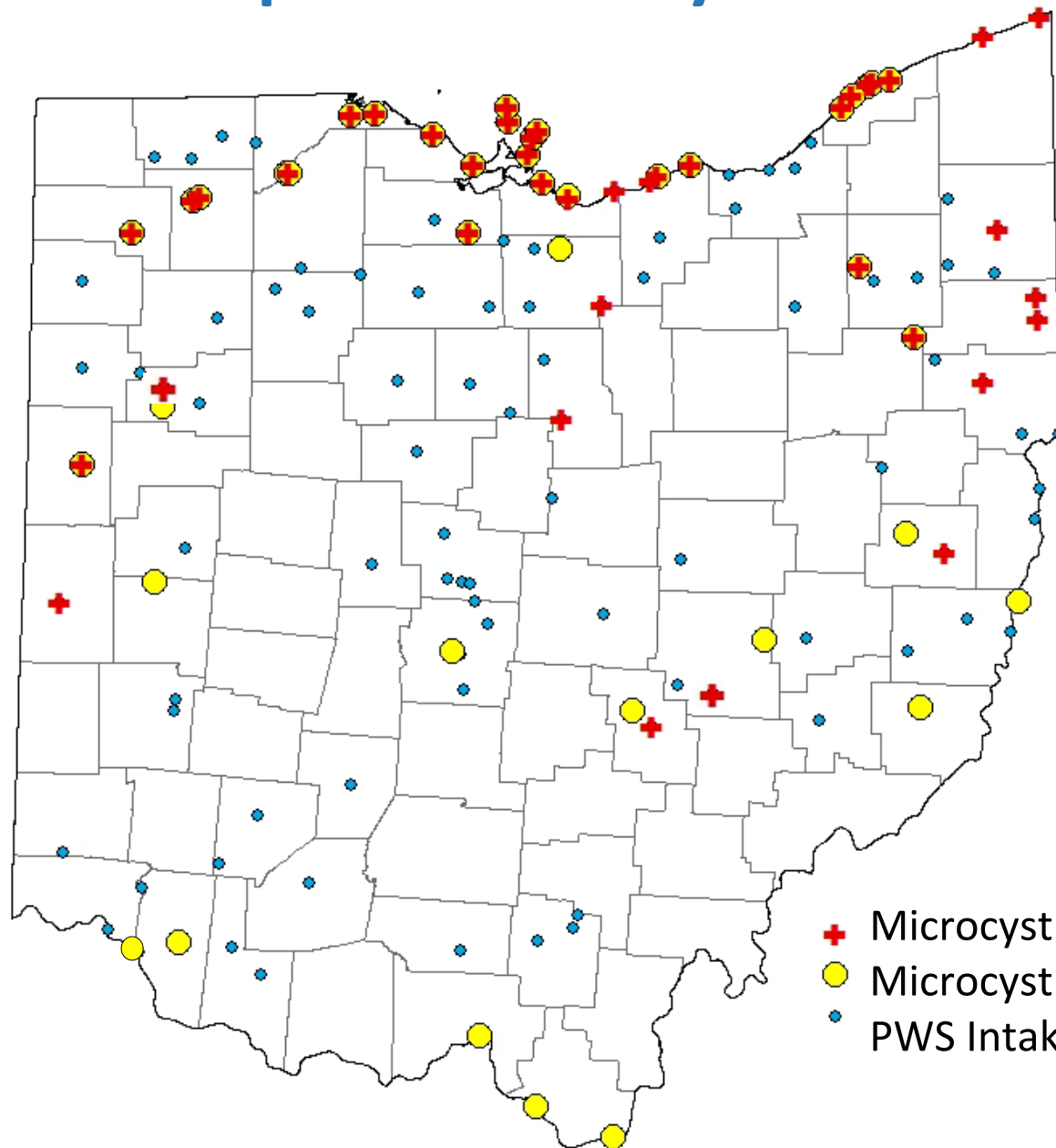
Raw Water Microcystins at an Inland PWS – Microcystins Do Not Always Peak in Summer



Late Fall-Winter Microcystins and qPCR Trends



Widespread Microcystins Occurrence in Ohio



Source Water
Microcystins
Detections at
Ohio PWSs

Data Available for 119
Public Water System
Source Waters

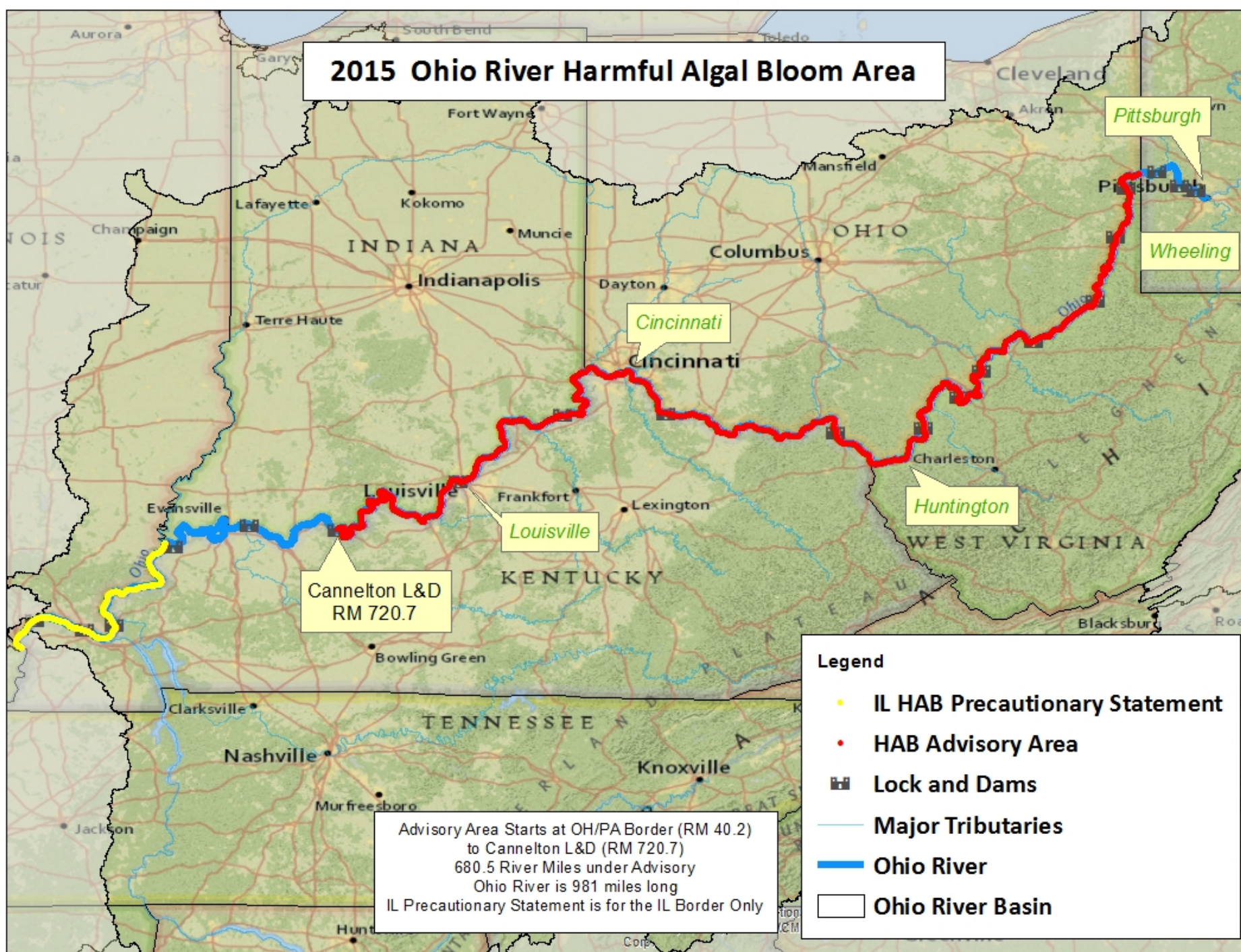
(includes ~2000
compliance samples
collected in 2016)

- + Microcystins Detections June –Dec. 2016
- Microcystins Detections 2010-2015
- PWS Intakes

Widespread Occurrence: Lake Erie, Inland Lakes, Upground Reservoirs, Quarries, and Rivers

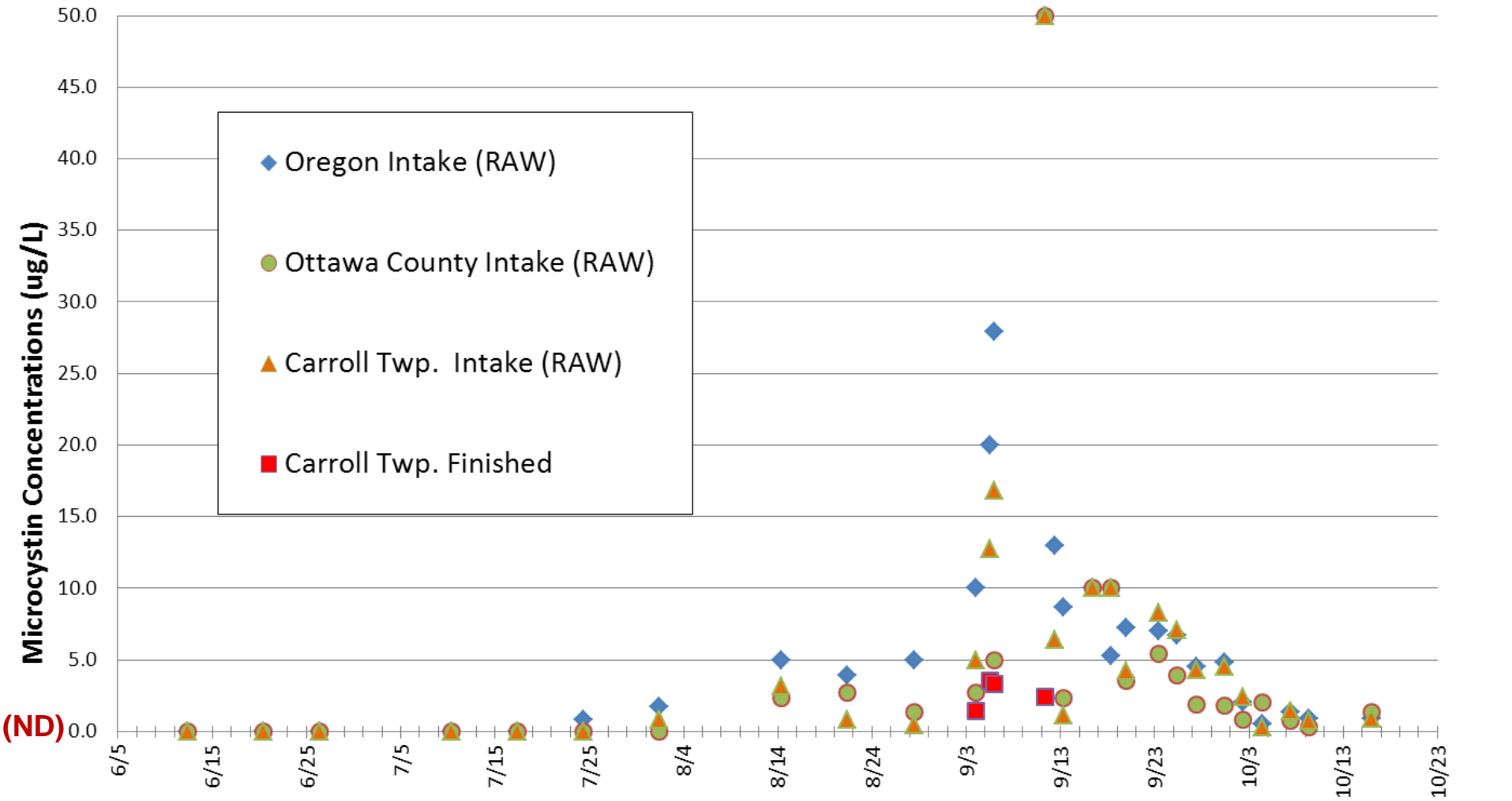


2015 Ohio River Harmful Algal Bloom Area



Microcystins Concentrations Can Increase Rapidly

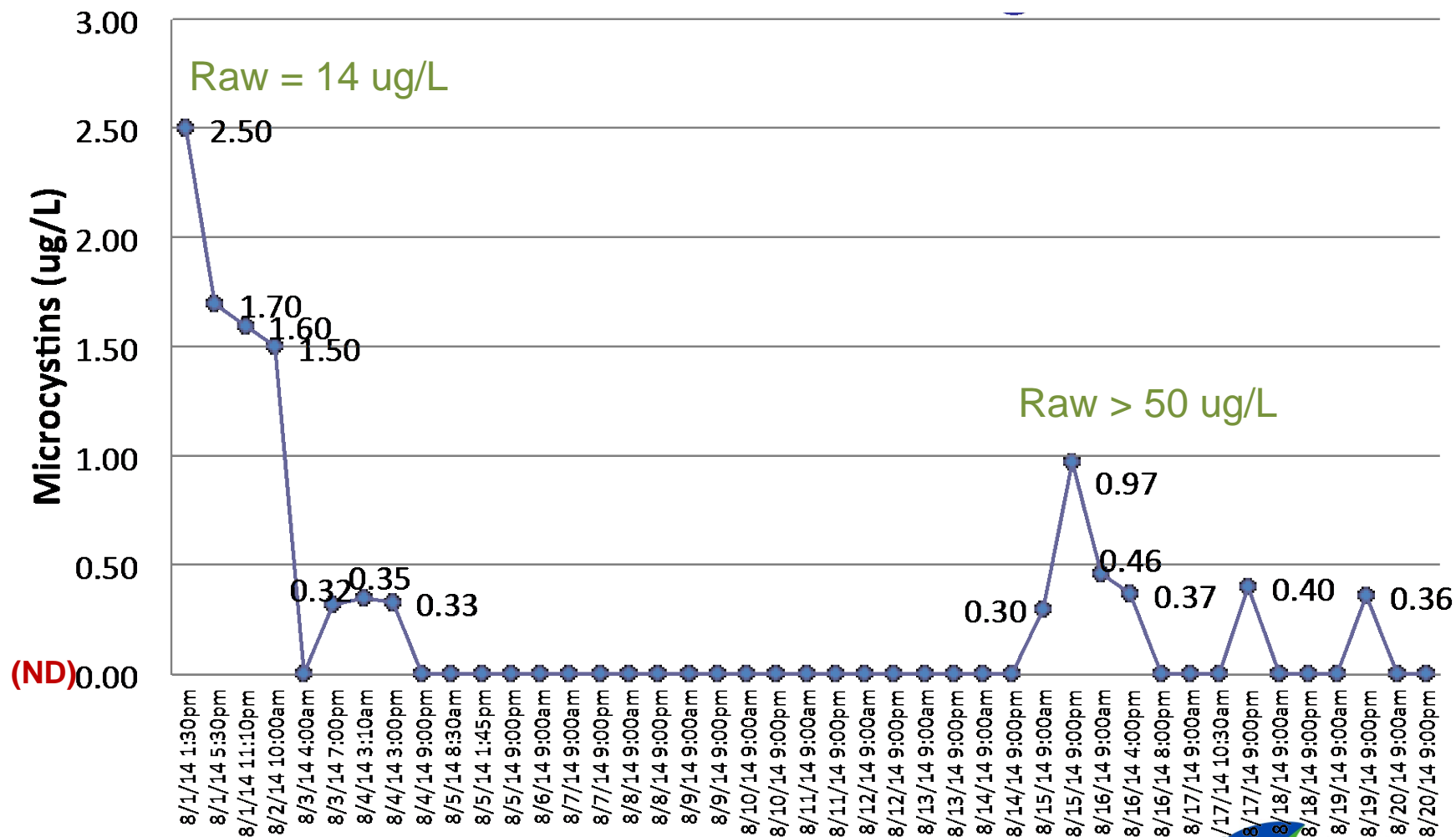
2013 Western Lake Erie Basin Microcystin Concentrations



ND= Not Detected (Concentration < 0.3)

Source: Oregon

Microcystin Source Concentration and Treatment Capacity (Finished Water Concentrations Lake Erie PWS)

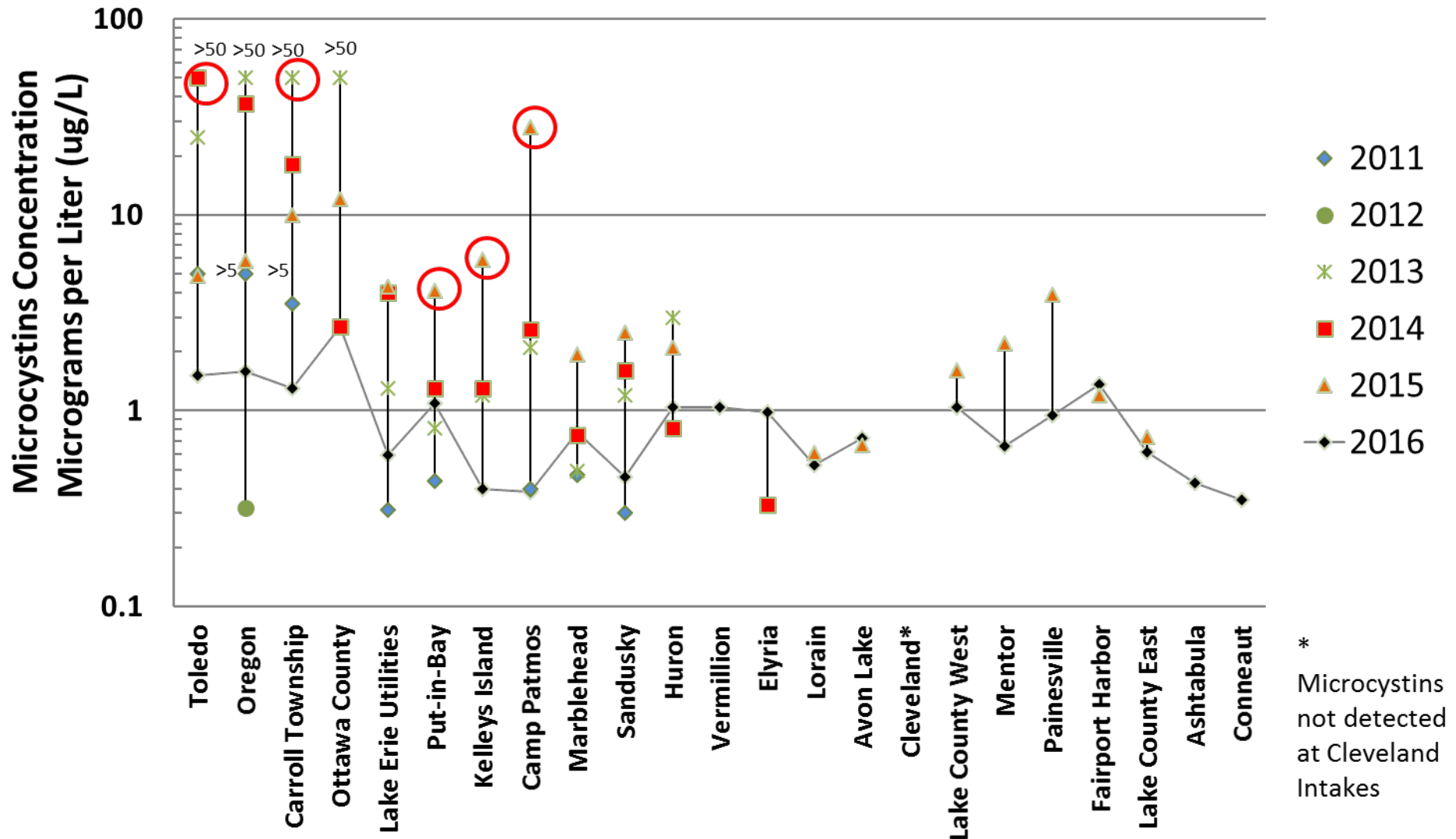


Microcystins also detected at 31
distribution sites

ND= Not Detected (Concentration <0.30)

HAB Severity Can Vary Year to Year

Maximum Microcystins Concentrations at Lake Erie Public Water System Intakes (2010 to 2016)



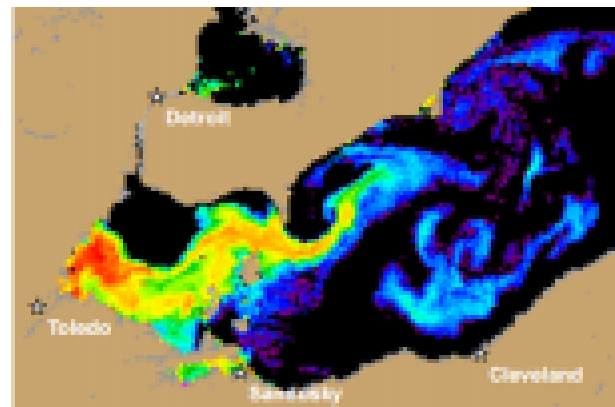
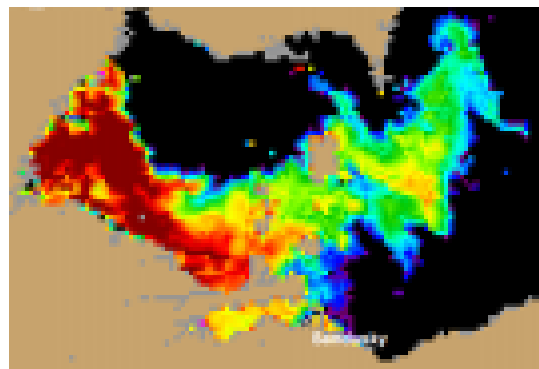
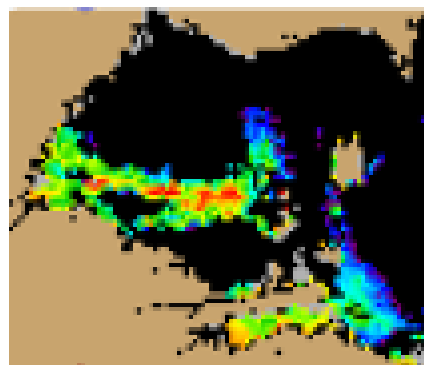
Lake Erie HAB Comparison 2015 and 2016

July

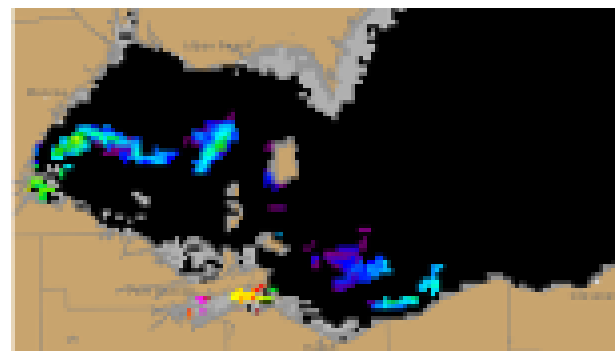
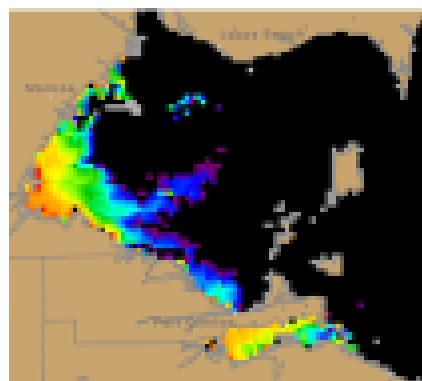
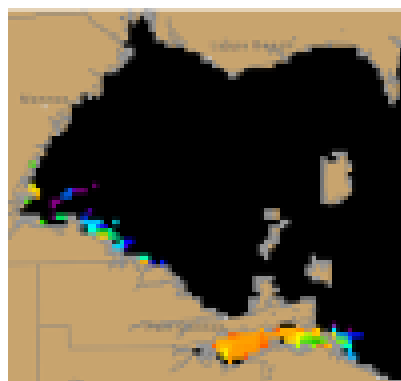
August

September

2015

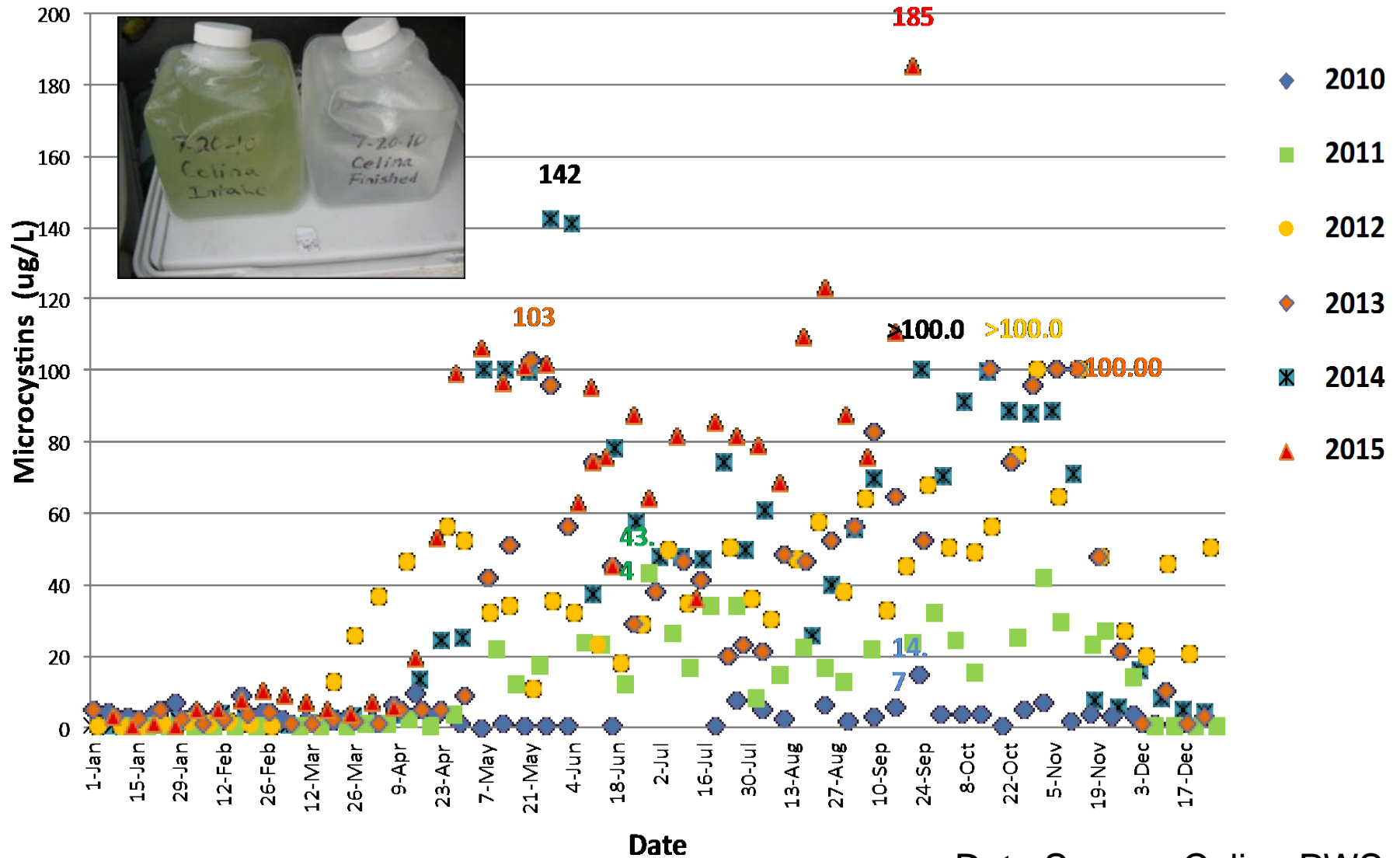


2016



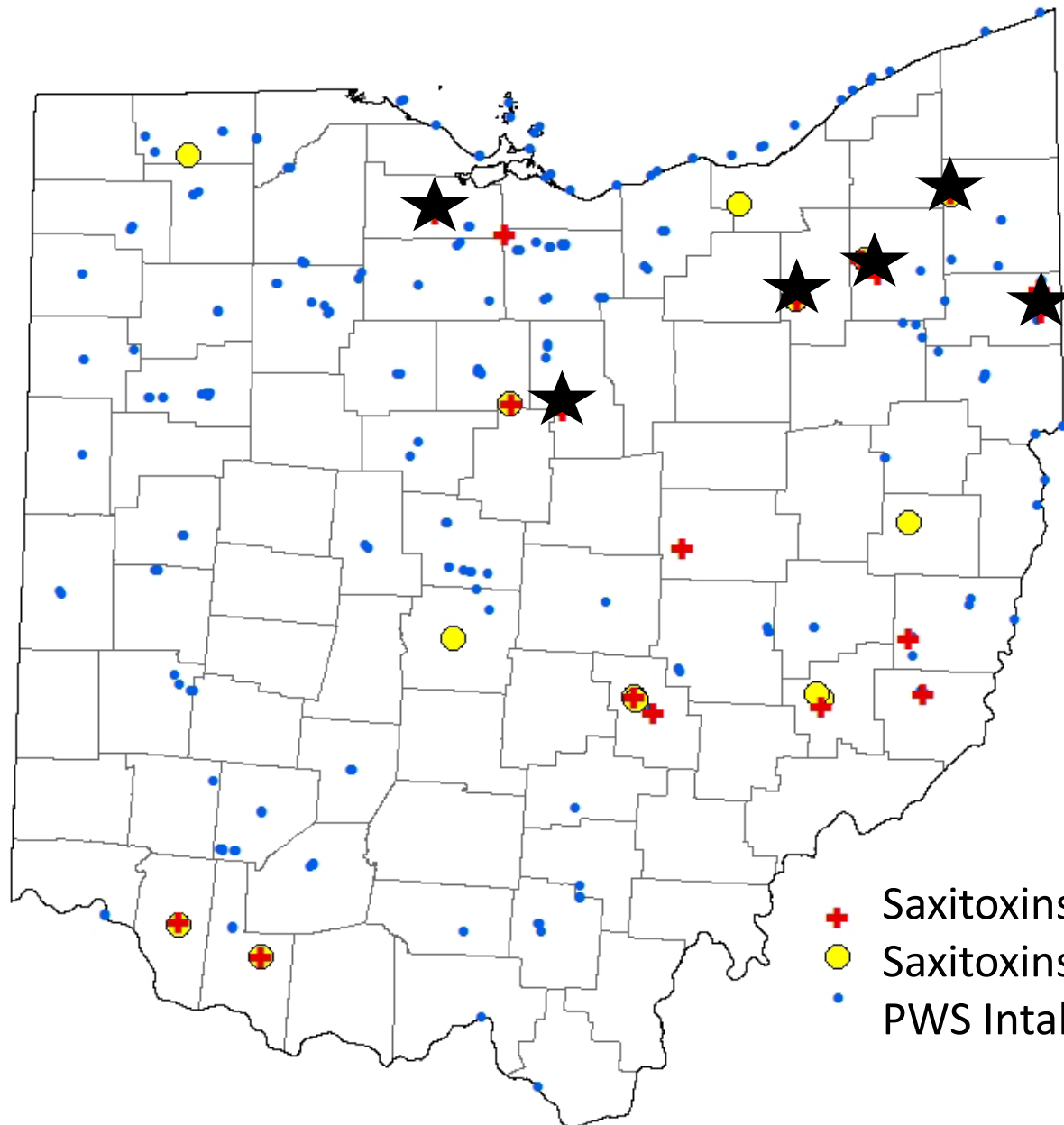
Was 2016 a good year to benchmark?

Grand Lake St. Marys Microcystin Concentrations at City of Celina Intake (Raw Water)



Data Source: Celina PWS

Saxitoxins are an Emerging Issue

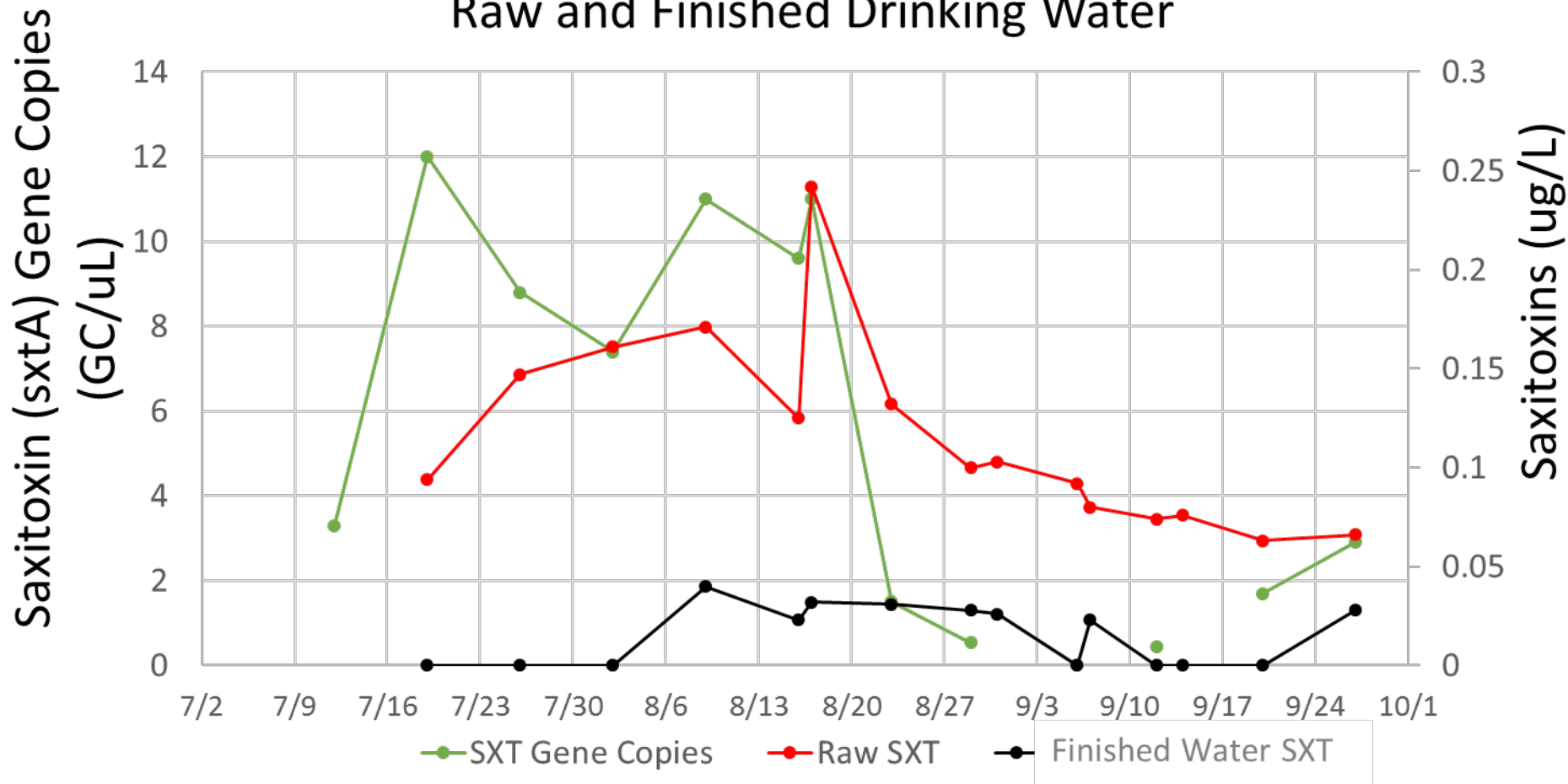


★ 2016 Finished
Water Saxitoxins
Detections
(below
threshold)

Benthic cyanobacteria
may be a source of
freshwater saxitoxins.

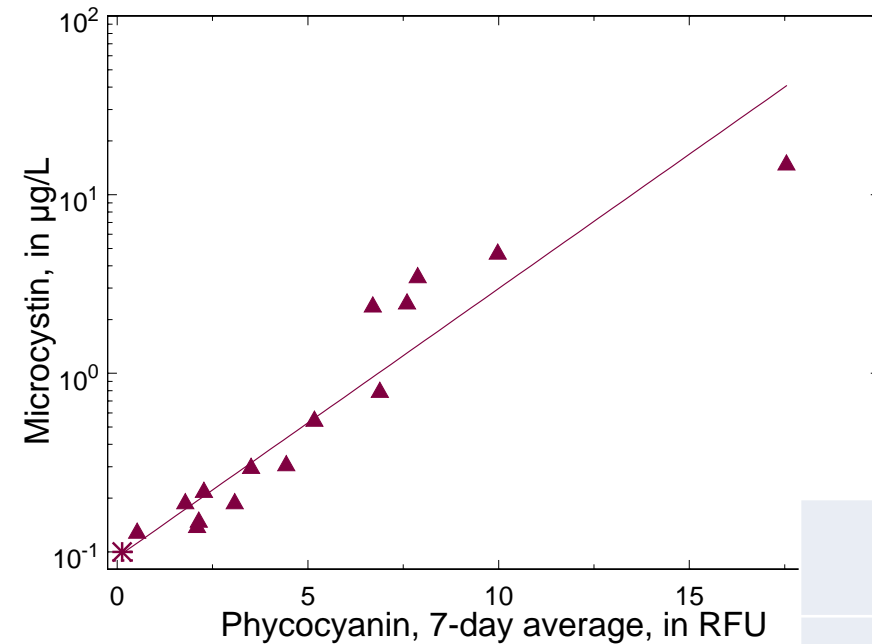
+ Saxitoxins Detections 2016
● Saxitoxins 2010-2015
● PWS Intakes

Saxitoxin Gene and Saxitoxins Detections in Raw and Finished Drinking Water



Multiplex Assay Works: Microcystin (mcyE) genes and microcystins also detected; Cylindrospermopsin (cyrA) genes also detected (cylindrospermopsin was not detected)

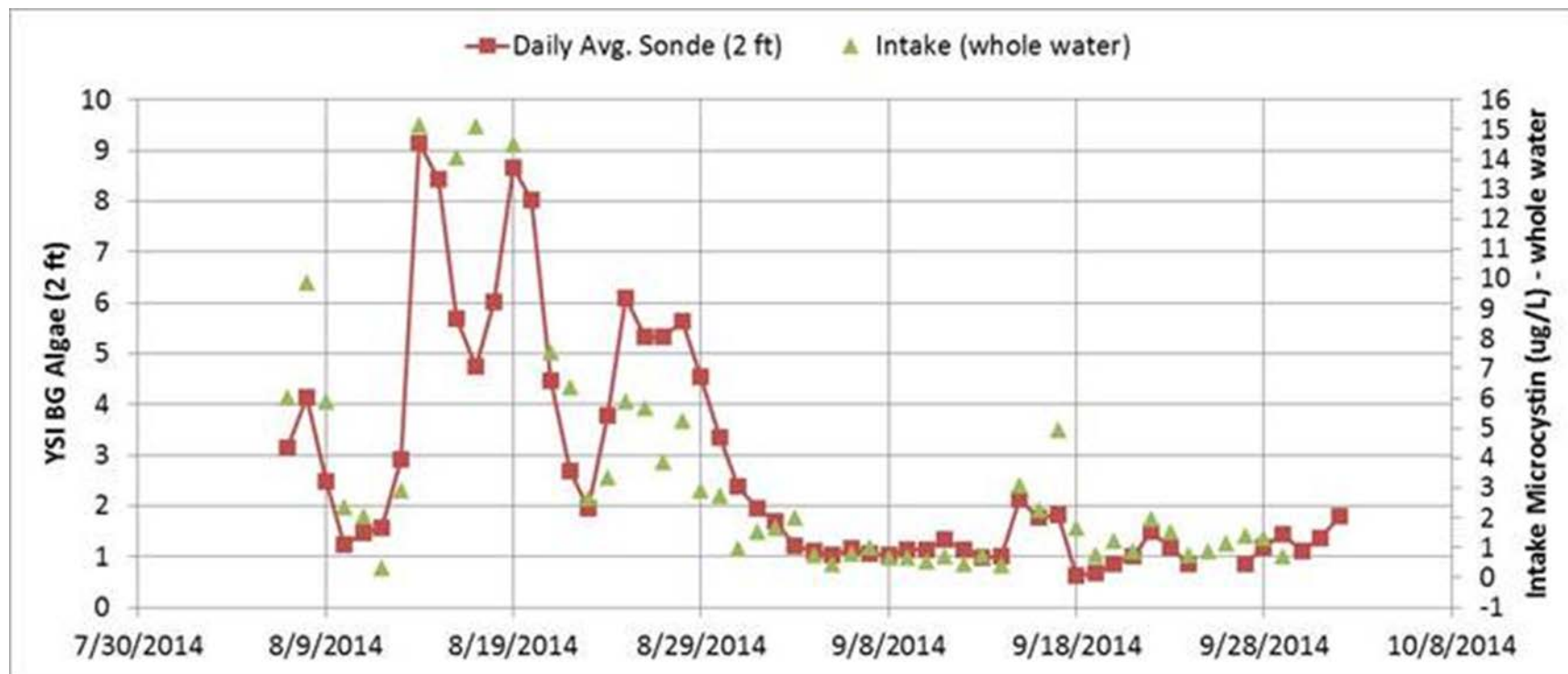
Use of Multi-Parameter Datasondes for Continuous Monitoring



Data Courtesy Donna
Francy, USGS

Spearman's correlation to microcystin concentrations	rho	p
Phycocyanin, 7-day average	0.98	<0.0001
Dissolved oxygen, 14-day average	0.88	<0.0001
pH, 7-day average	0.83	<0.0001
Temperature, instantaneous 10 a.m.	0.73	0.0031
Chlorophyll, 24-hour average	0.53	0.0358
Specific conductance, 3-day average	-0.20	0.4473

Microcystin Concentration and Phycocyanin Fluorescence at Toledo's Intake



-Graph provided to Ohio EPA by Ed Verhamme, Limnotech.

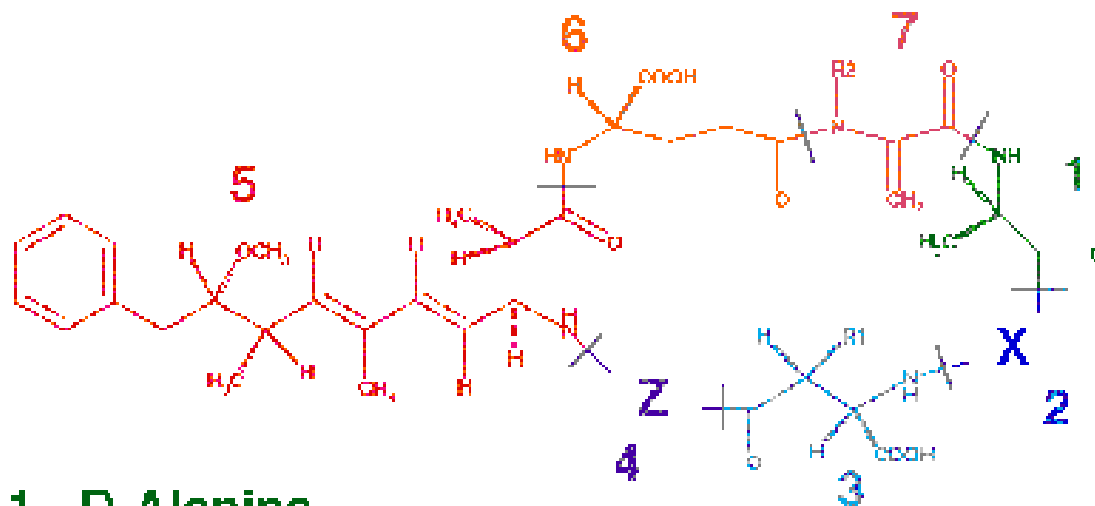
Ohio EPA provided funding to 36 PWSs for purchase of phycocyanin sondes or readers

Phycocyanin Data Interpretation

- Phycocyanin Concentrations vary based on type of cyanobacteria present, turbidity of the water and other factors.
- Relative/Raw Fluorescence Units (RFUs) better than Cell Counts.
 - Can calibrate to cell counts in source water, but this can change if cyanobacteria genera shift or turbidity changes.
- Evaluate trends, not absolute values.

Lessons Learned – Analytical Methods Matter

No “Perfect” Analytical Method for Detecting TOTAL Microcystins



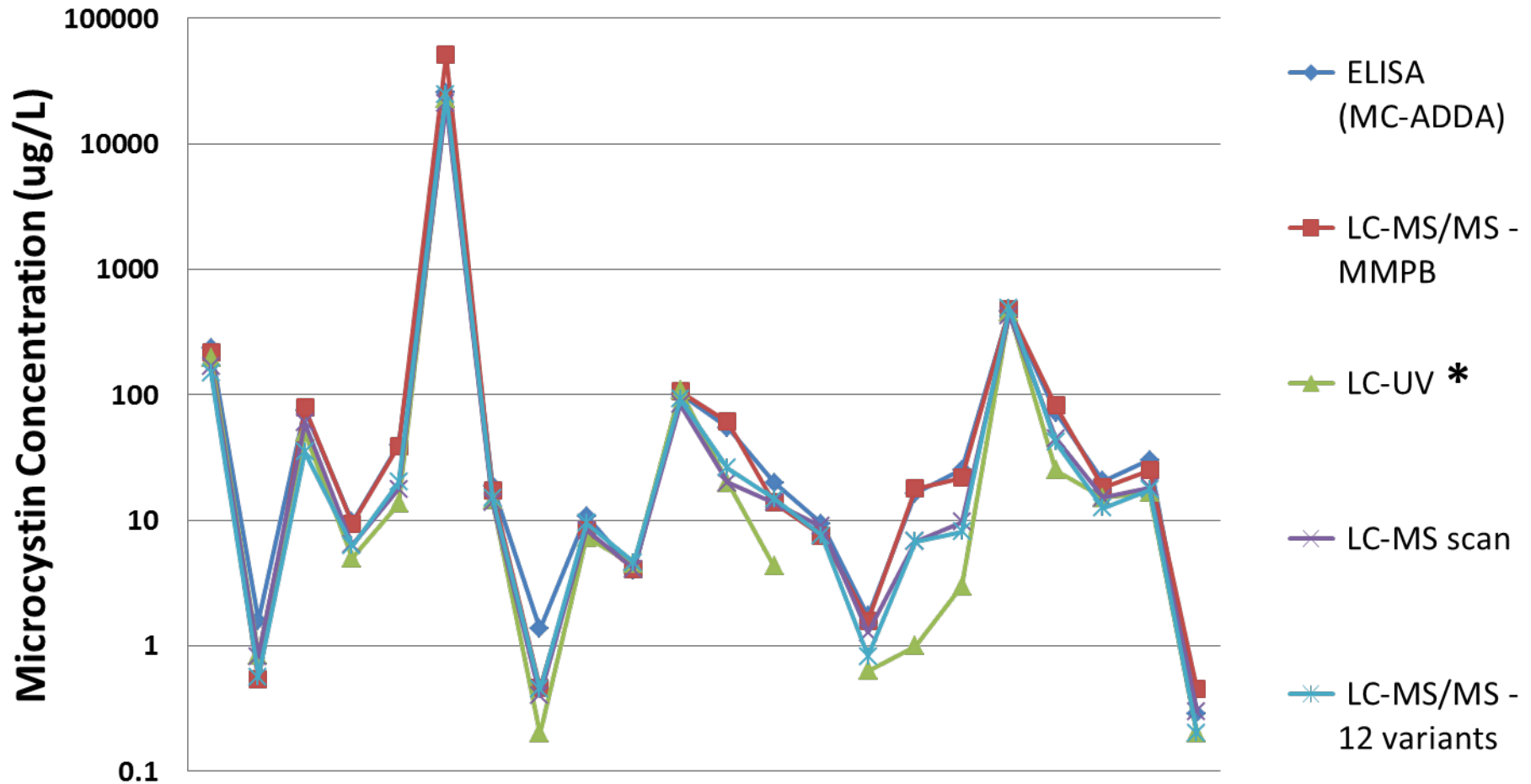
- 1 - D-Alanine
- 2 - Variable L-amino acid
- 3 - D-Methylaspartic acid
- 4 - Variable L-amino acid
- 5 - 3-amino-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoic acid (Adda)
- 6 - D-Glutamic acid
- 7 - N-Methyldehydroalanine

- Over 140 Microcystin Variants
- Standards Not Available for Majority

Analytical Method Comparison & Microcystin Variant Evaluation

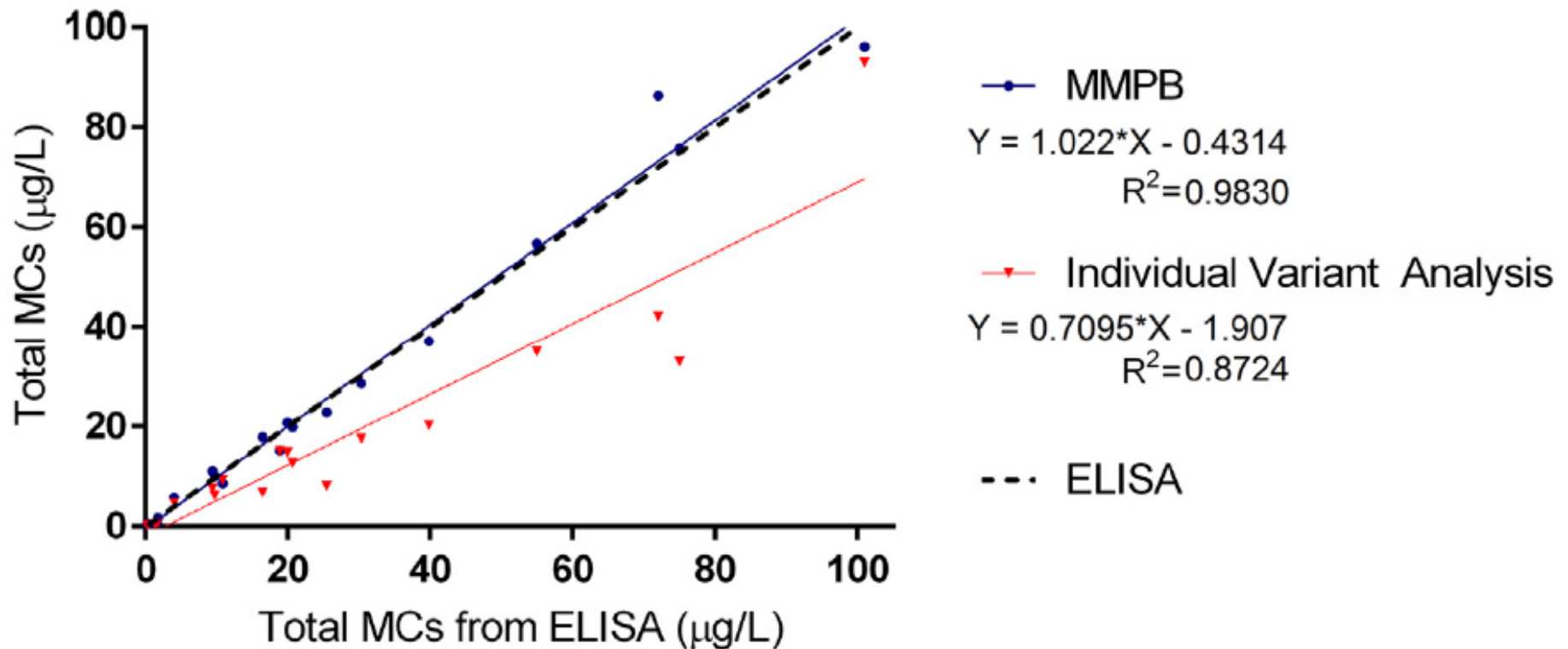
- 11 Sites: 4 up-ground reservoirs, 2 in-stream reservoirs, 2 Lake Erie locations, 2 canal-feeder lakes, and 1 river source
- 22 samples from 2014 selected to help evaluate spatial and temporal variability within source waters
- Variety of cyanobacteria genera represented
- Each sample analyzed using 5 separate analytical methods ELISA-MC-ADDA, LC-UV/PDA, LC-MS/MS – individual variant (12), LC-MS/MS-MMPB, LC-MS scan

Results of Method Comparison



* LC-UV data presented do not include false-positives that were eliminated from total (Based on lack of confirmation with LC-MS methods). Sample #14 was non-detect for MCs using LC-UV.

Results of LC-MS/MS MMPB and Individual Variant Analysis Compared to ELISA



Microcystins Methods - Key Findings

- 16 different MC-variants detected
- MC-LR was only detected at 5 of 11 sites (45%)
- Most commonly detected variants were: MC-YR, [Dha7] MC-LR and [DAsp3] MC-RR
- LC-PDA methods prone to interference, potential for false positives and false negatives
- LC-MS/MS MMPB method confirmed ELISA results (raw water)
- 91%-100% of samples had MC-variants not detectable by U.S. EPA Method 544 (including dominant MC-variant in some samples)
- LC-MS/MS individual variant analysis under-reported total microcystins, based on MMPB, LC-UV/MS scans, and ELISA data
- No perfect method for TOTAL Microcystins

Analytical Methods Utilized by Ohio EPA

	Microcystins (µg/L)	Cylindro- spermopsin (µg/L)	Saxitoxins (µg/L)	Anatoxin-a (µg/L)
Surveillance sampling	ELISA (MC-ADDA)	ELISA	ELISA	LC-MS/MS
Repeat sampling in response to a finished water detection	ELISA (MC-ADDA)	LC-MS/MS	LC-MS/MS	LC-MS/MS

ELISA: Enzyme-Linked Immunosorbent Assay

LC-MS/MS: Liquid Chromatography followed by tandem
Mass Spectrometry



Ohio HAB Initiatives & Resources

PWS Funding: \$1.5 million in grants for analytical kits, microscopes, and raw water quality sensors and \$100 million in zero-interest loans for drinking water infrastructure improvements including additional treatment, alternative sources and storage.

Technical Assistance: Site visits with all susceptible surface water systems on treatment optimization and contingency planning. Numerous presentations, webinars and workshops have been held with PWSs, stakeholders, and partners.

Guidance: Developed Cyanotoxin Treatment White Paper, Cyanotoxin Treatment Optimization Protocol, Cyanotoxin General Plan, Algaecide Application Fact Sheet, Distribution Cyanotoxin Monitoring; ELISA SOPs, Response Strategies; Website.

Emergency Response: Established PWS-specific HAB incident action plans and coordinated with partner agencies during multiple table top exercises and drills.

Clean Water Act Integration: Established cyanotoxin indicators of impairment for determining public drinking water supply beneficial use attainment

Research: Partnered with university and government researchers to address data gaps.

www.epa.ohio.gov/ddagw/HAB.aspx



Public Drinking Water Supply Beneficial Use

First Listings in 2014 IR

Algae Impairment Criteria

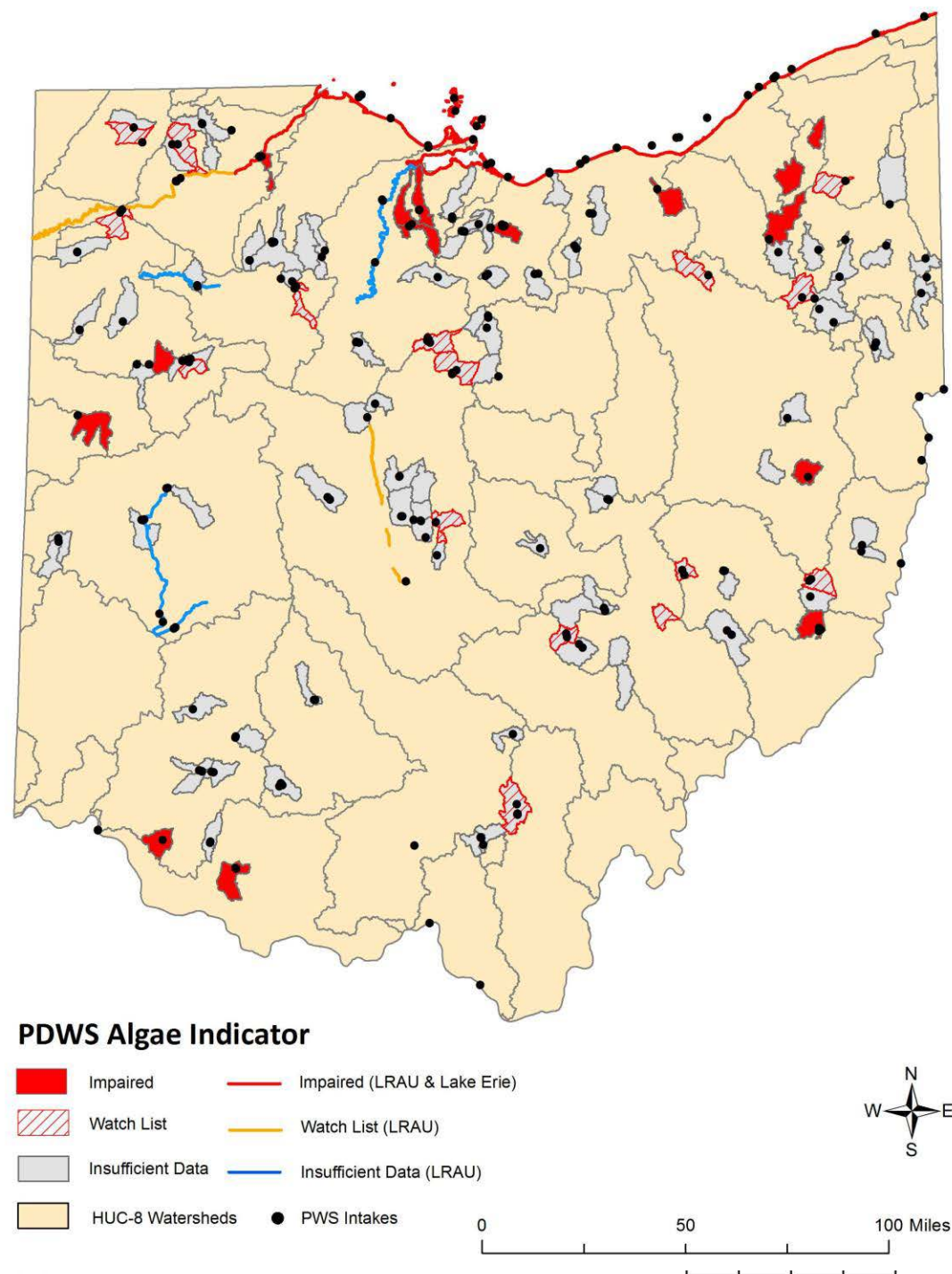
2016 CWA Integrated Water Quality Monitoring and Assessment Report:

Impaired = at least 2 source water cyanotoxin detections above drinking water thresholds at least 30 days apart.

22 Public Water Systems Triggered Impairment Listings

The Entire Lake Erie Basin Shoreline and 15 Watersheds are Impaired

19 additional watersheds are on a Watch List



Summary

- HABs are Not Always Visually Apparent
- HABs Can Occur Year-Round, in a Variety of Source Waters, and can be Different (or similar) Every Year
- Saxitoxins are an Emerging Issue
- Early Warning Monitoring Helps
- Methods Matter
- Plan Ahead!

Questions?

www.epa.ohio.gov/ddagw/HAB.aspx

Heather.Raymond@epa.ohio.gov

(614) 644-2752

