

Sampling Protocols

Barry H. Rosen, Ph. D.

Office of the Southeast Regional Director
(CFLWSC) Orlando, FL

brosen@usgs.gov

407-803-5508

Recognizing a cyanobacteria bloom: field images (blue-green to greenish in color)



Field and Laboratory Guide to Freshwater Cyanobacteria Harmful Algal Blooms for Native American and Alaska Native Communities



Open-File Report 2015–1164

U.S. Department of the Interior
U.S. Geological Survey

brosen@usgs.gov

Getting a Sample qualitative

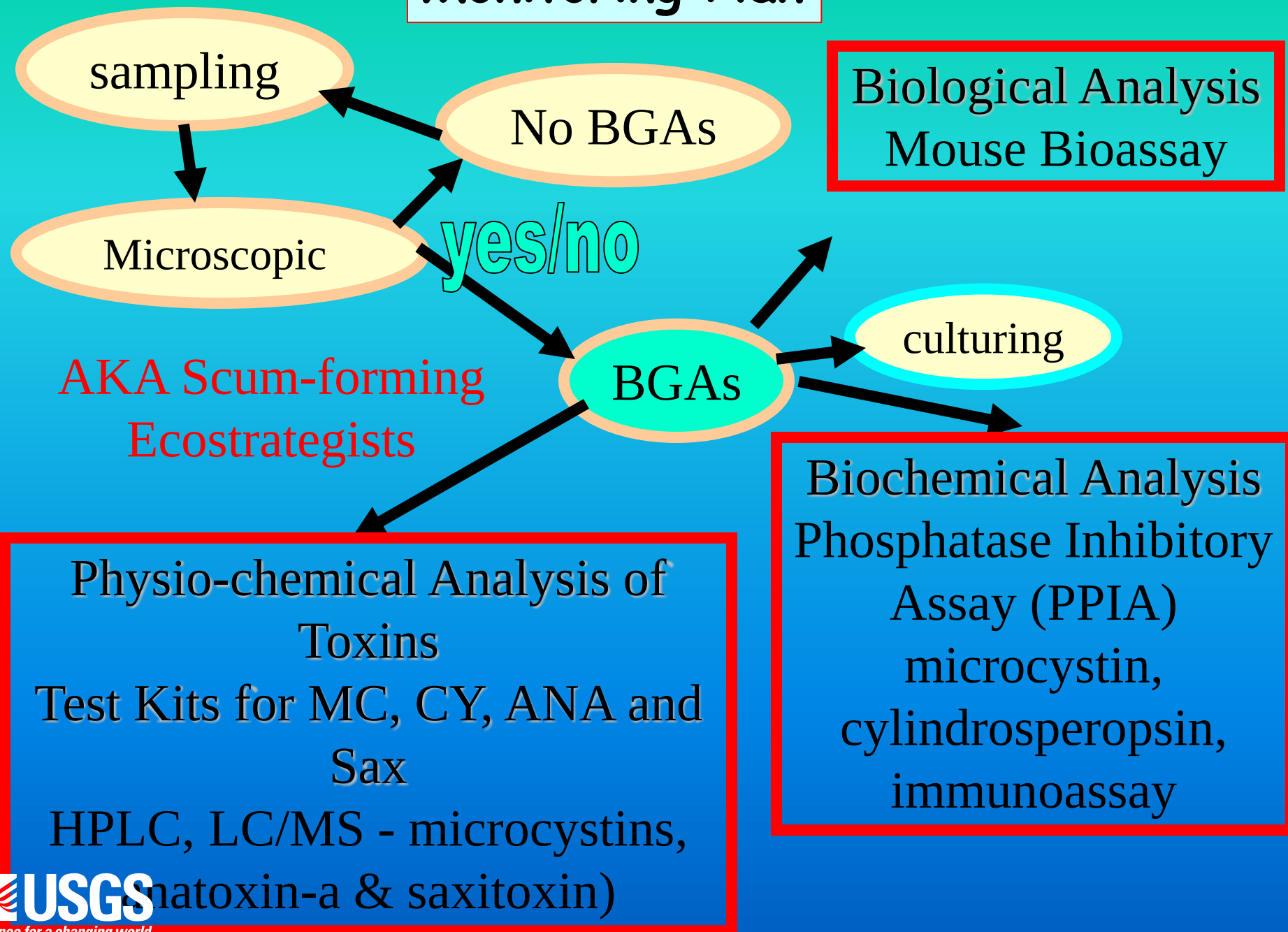


Dense: use a glove!



**Not dense: use a
plankton net!**

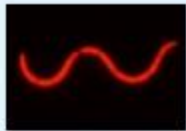
Monitoring Plan



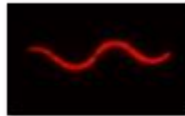
Probes



Qualitative: Diversity and Potential toxin producers



Cylindrospermopsis Mattamuskeet W. 10-22-15 WG curvy 2 cyano



Cylindrospermopsis Mattamuskeet W. 10-22-15 WG curvy cyano



Komvophoron 1 Mattamuskeet E. 7-22-15 Live 930 am cyano



Komvophoron 2 Mattamuskeet E. 7-22-15 930 am cyano



Komvophoron Mattamuskeet E. 9-10-15 Live 900 am cyano



Komvophoron Mattamuskeet W. 7 22 15 K. apiculatum me...



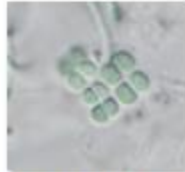
Komvophoron Mattamuskeet W. 7-22-15, culture 8-3-15 cyano



Komvophoron Mattamuskeet W. 7-22-15, culture 8-7-15 1a cyano



Merismopedia Mattamuskeet E. 7-22-15 Live 930 am cyano



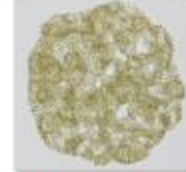
Merismopedia Mattamuskeet W. 7-22-15, culture 8-3-15 cyano



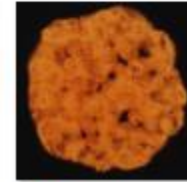
Merismopedia Mattamuskeet E. 7-22-15 930 am cyano



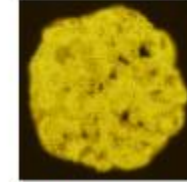
Merismopedia Mattamuskeet W. 7-22-15, culture 8-7-15 cyano



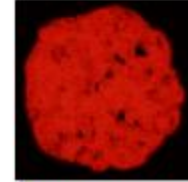
Microcystis culture 1a LM M. aeruginosa cyano



Microcystis culture 1a UV M. aeruginosa cyano



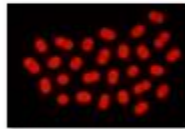
Microcystis culture 1a WB M. aeruginosa cyano



Microcystis culture 1a WG M. aeruginosa cyano



Microcystis Mattamuskeet W. 10-22-15 LM mea M. aeruginosa c...



Microcystis Mattamuskeet W. 10-22-15 WG M. aeruginosa cyano



Microcystis Mattamuskeet W. 10 22 15 M. aeruginosa cyano



Microcystis Mattamuskeet W. 10 22 15 mea M. aeruginosa cyano



Phormidium Mattamuskeet E. 8 13 15 1a cyano



Phormidium Mattamuskeet E. 8 13 15 1b cyano



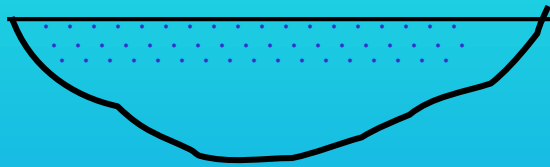
Phormidium Mattamuskeet E. 8 13 15 c cyano



Phormidium Mattamuskeet E. 8 13 15 c mea cyano

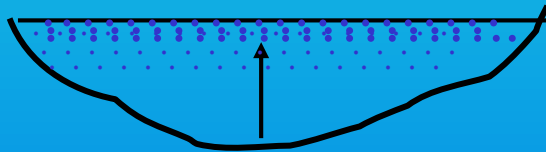
Beware of this phenomenon

initial distribution



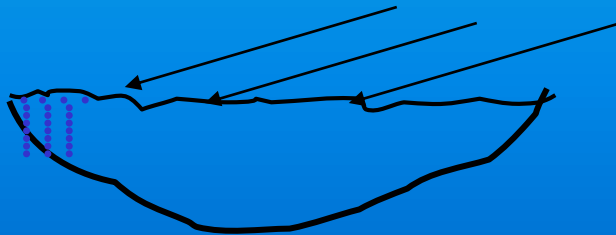
**100,000 cells/L;
20 $\mu\text{g/L}$ toxin**

buoyancy



**10,000,000 cells/L;
2000 $\mu\text{g/L}$ toxin**

wind

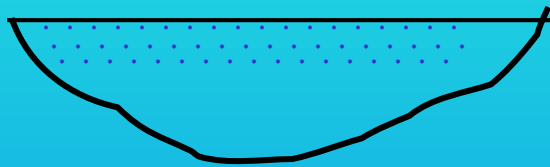


**100,000,000 cells/L;
20,000 $\mu\text{g/L}$ toxin**

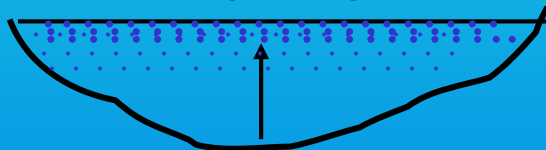
5,000-11,600 $\mu\text{g/kg}$ bw causes liver damage = 2 mg in 10 kg child

Where do I sample?

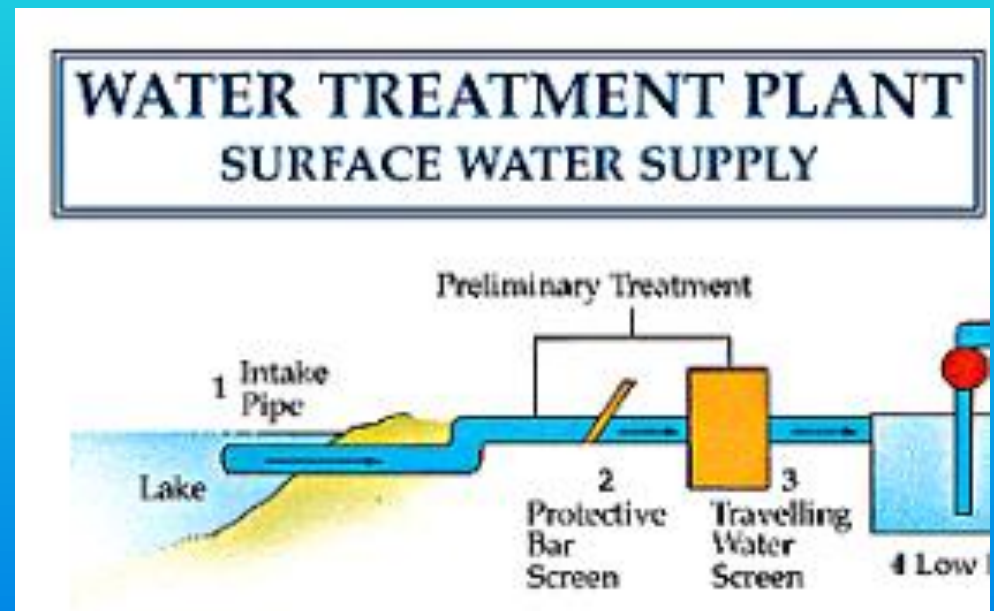
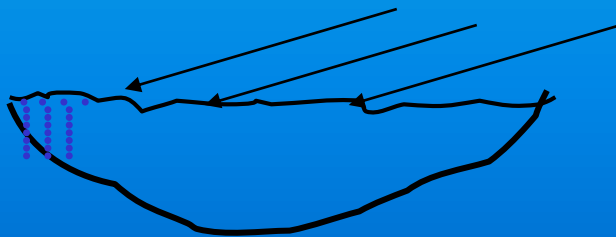
initial distribution



buoyancy



wind



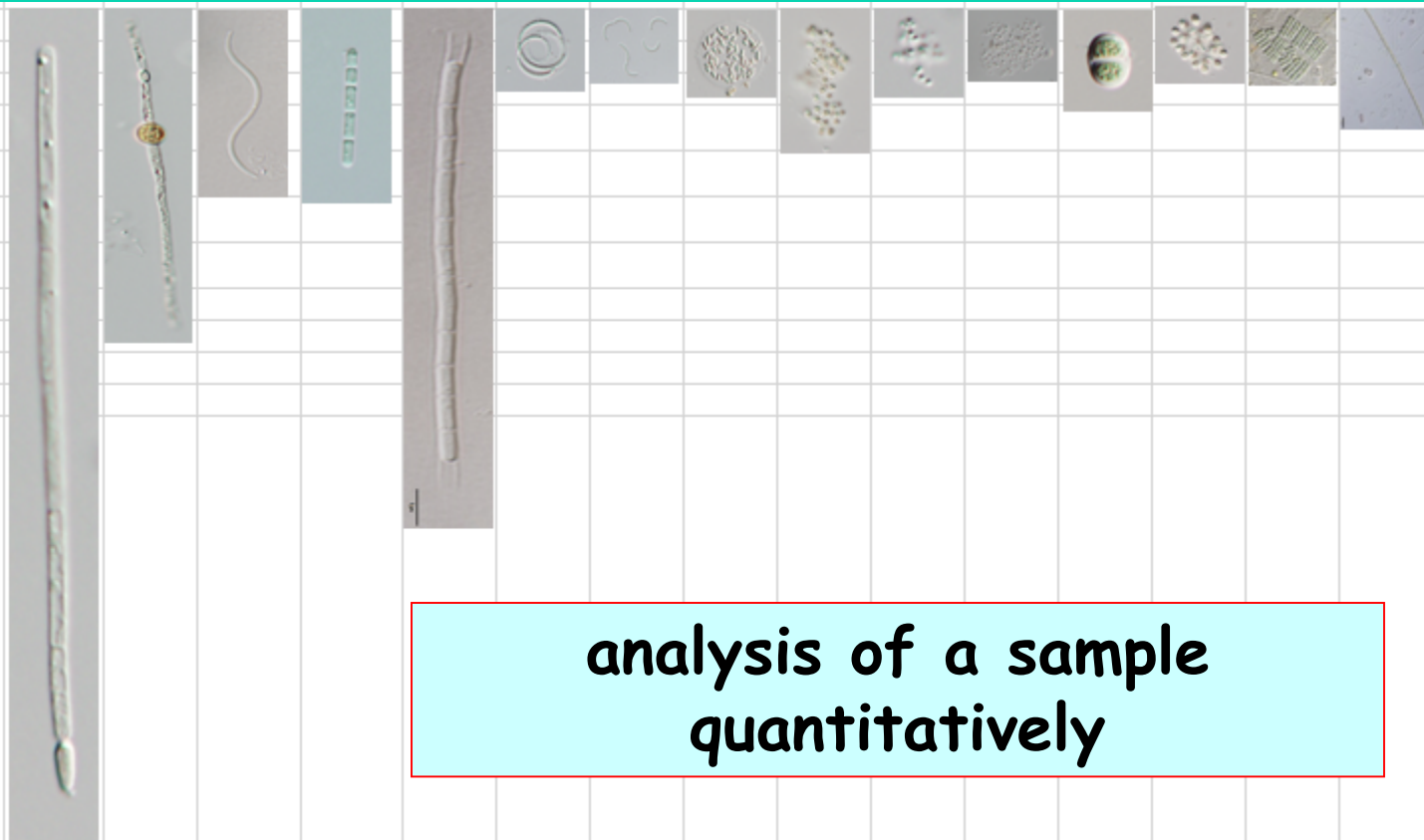
Getting a Sample quantitative



Van Dorn

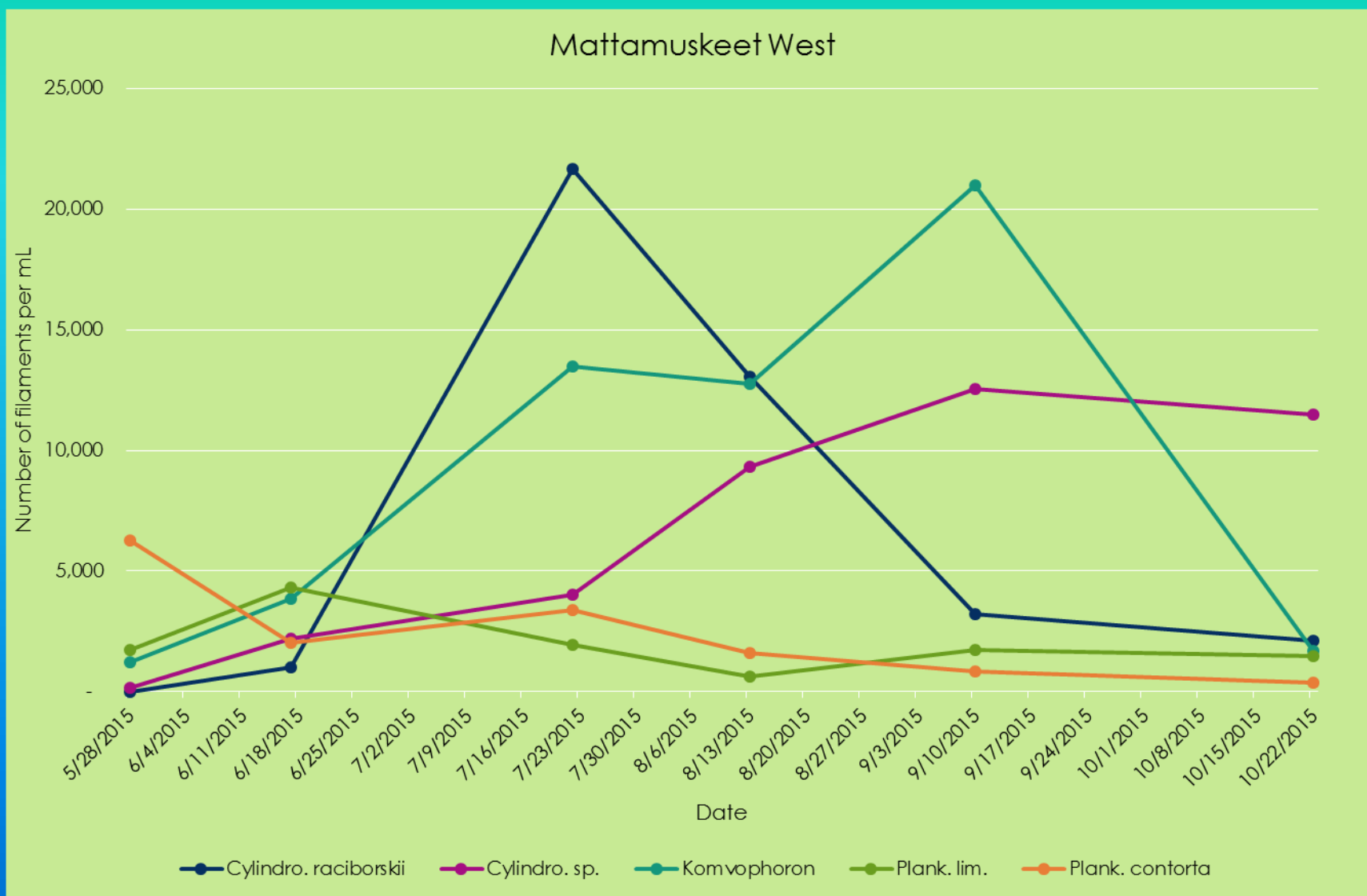


Depth Integrated
Sampling

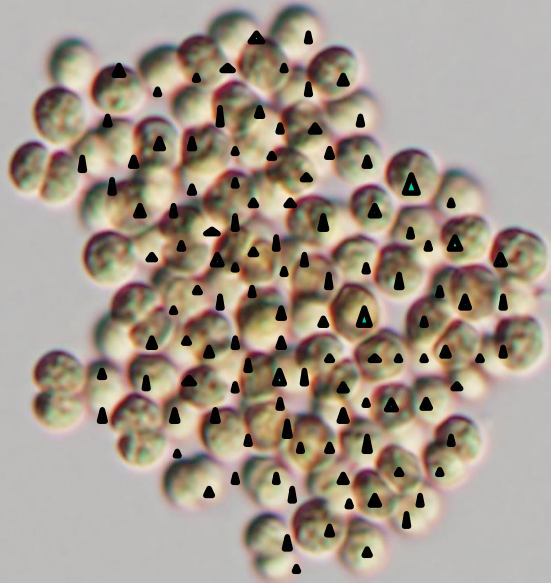


analysis of a sample quantitatively

				<i>Cylindrocapsa raciborskii</i>	<i>Chrysosporum ovalis</i>	<i>Cylindrocapsa cf. catenella</i>	<i>Komvophoron</i> sp.	<i>Planktothrix limnetica</i>	<i>Planktolyngbya contorta</i>	<i>Romeria</i> sp.	<i>Aphanocapsa stagnina</i>	<i>Microcystis aeruginosa</i>	<i>Aphanocapsa nibilum</i>	<i>Aphanocapsa</i> sp.	<i>Chroococcus</i> sp.	<i>Snowella</i> sp.	<i>Merismopedia</i> sp.	<i>Sphaerospermopsis aphanizomenoides</i>
plankton/peri	Concentration factor	number of fields	cell count															
plankton	10x	6	311	0	0	4	3	44	127	0	3	1	3	69	0	2	28	0
plankton	10x	6	318	0	0	3	23	32	117	0	3	0	0	71	6	20	14	1
plankton	10x	7	307	9	5	33	17	67	53	1	1	0	0	59	6	14	8	0
plankton	10x	6	343	19	1	41	72	81	38	2	14	1	0	40	0	11	6	0

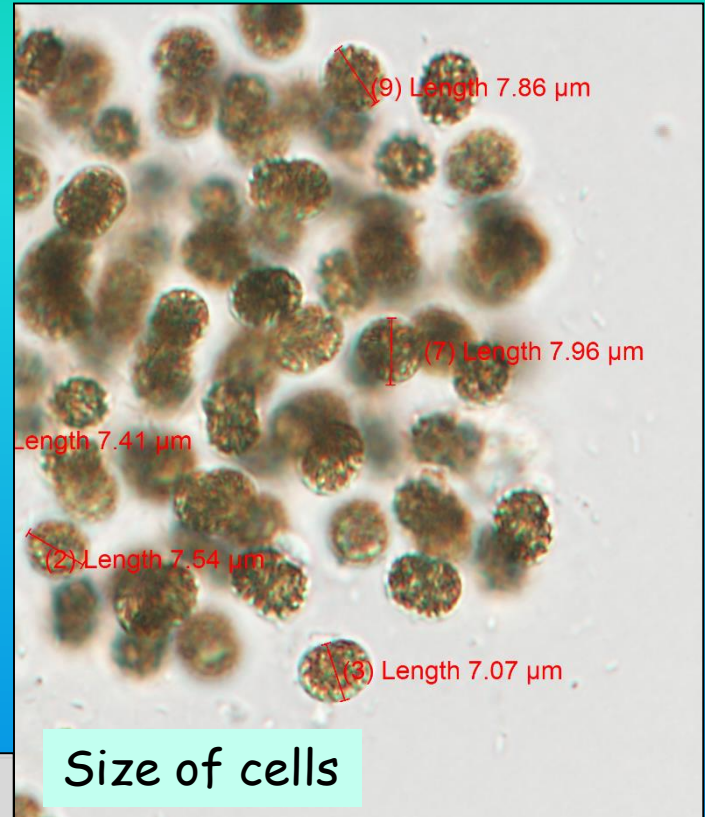


Biovolume per mL



Number of cells

5 μm



Size of cells

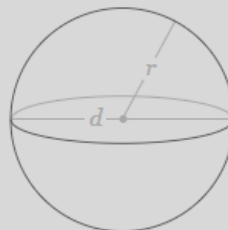
Sphere

Solve for volume ▾

$$V = \frac{4}{3} \pi r^3$$

r Radius

Enter value



How much of a sample and how should I “save” it

1. Collect 100 mL sample of a bloom live

Possible Methods:

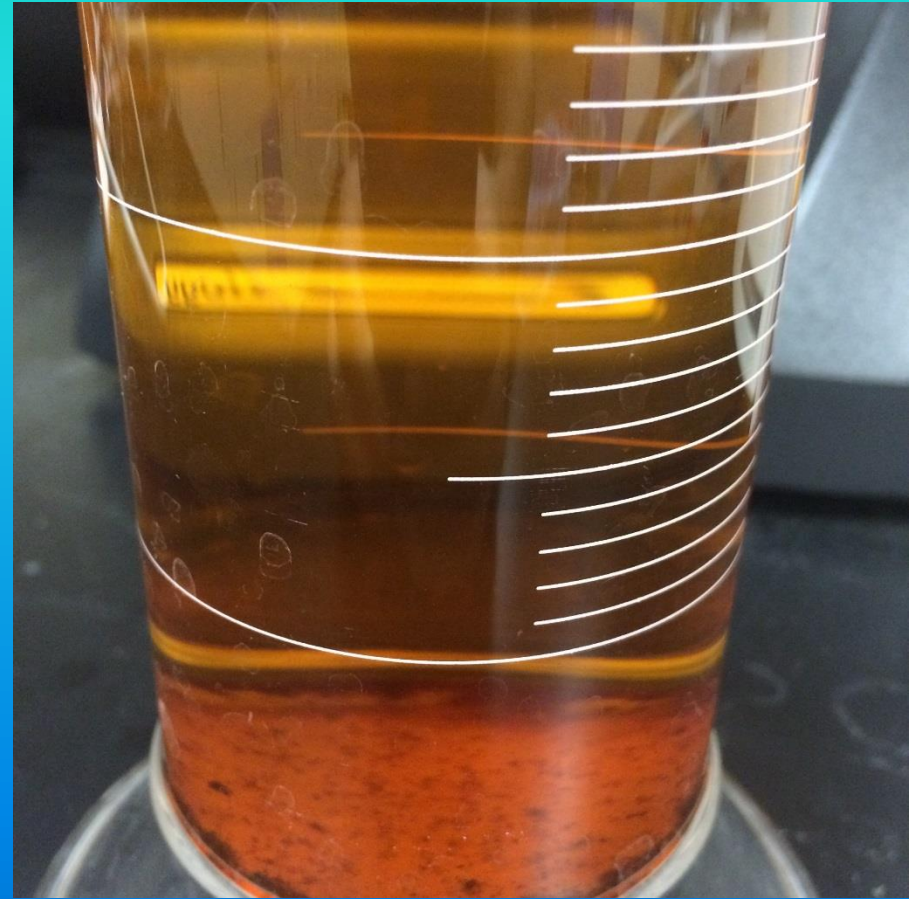
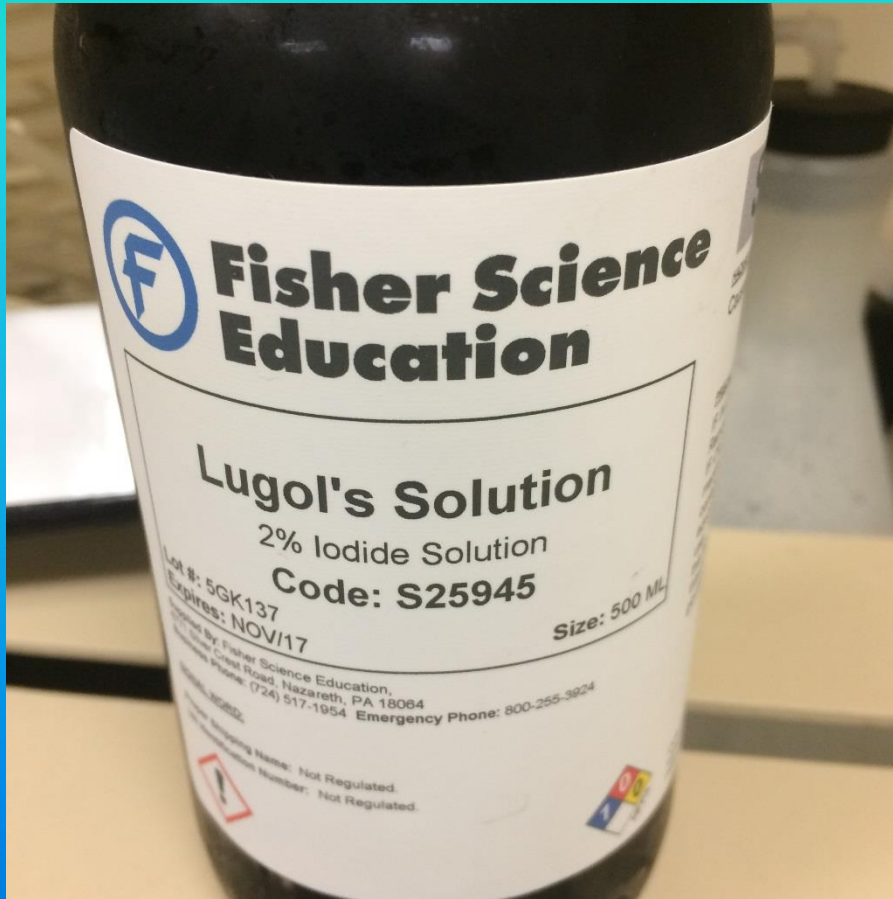
- a) A whole water sample by simple immersing a 500 mL bottle (glass or plastic) into a waterbody. The small volume in a large bottle allows for ample gas exchange during shipping.
- b) A plankton tow of a bloom, which concentrates a sample, and a liquid volume of 10 mL in a 100 mL bottle.

How much of a sample and how should I “save” it

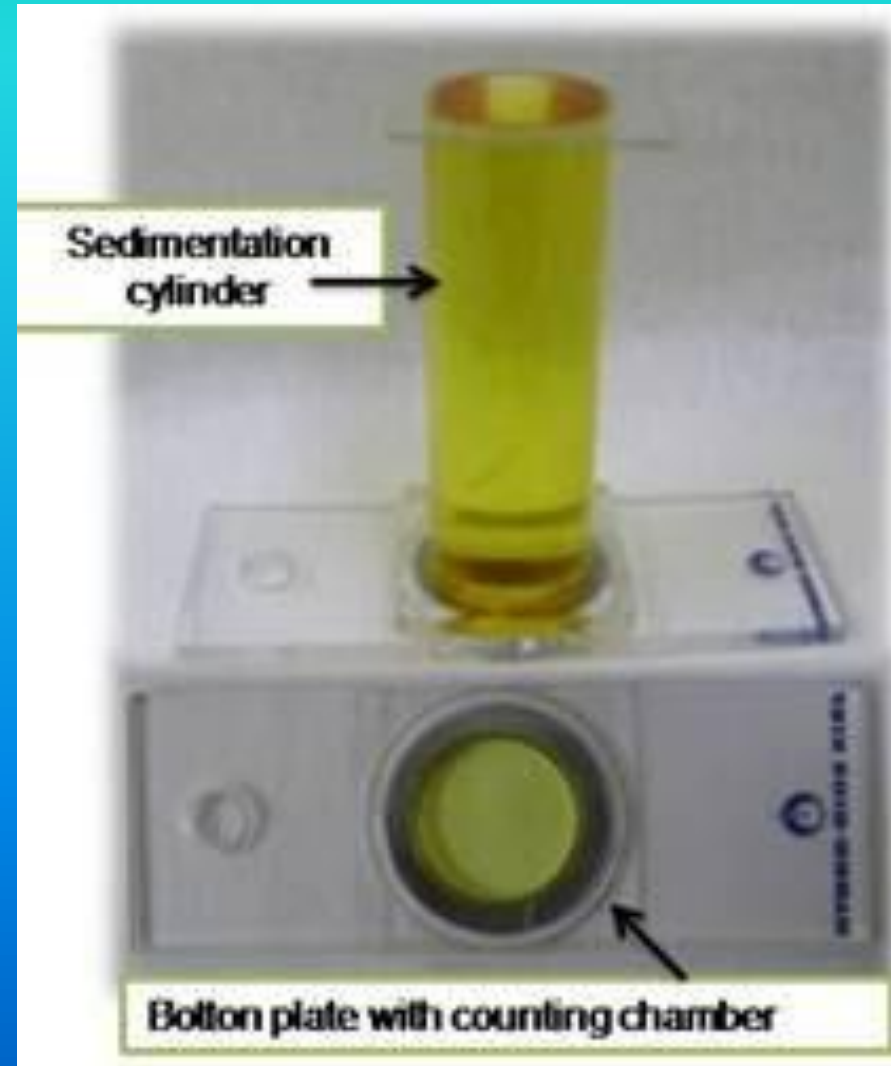
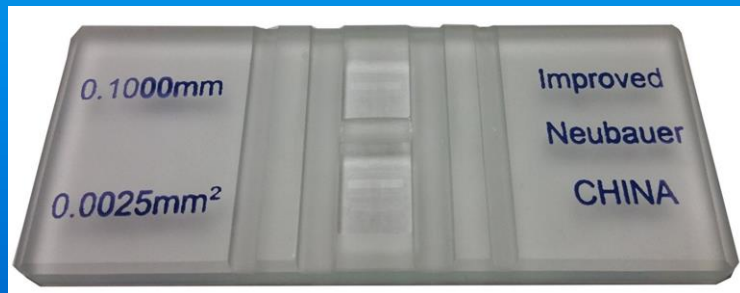
- 2) Collect 100 mL sample of a bloom, preserved with Lugol's iodine
- a) same procedures as step 1 to collect the samples
 - b) add 5% solution of Lugol's to turn the sample the color of tea.
(5% (wt/v) iodine (I_2) and 10% (wt/v) potassium iodide (KI) mixed in distilled water and has a total iodine content of 126.5 mg/mL).
-alternatively, Povidone-iodine can be used.



Lugol's and Settling



Quantitative Counting



Semi-Quantitative Counting and Toxins



Lab Instruments



Lab Instruments



How much of a sample and how should I “save” it for toxin analyses

3) Collect 1000 mL sample of a bloom, freeze it!



Cyanotoxins in Raw Water Sample Collection Quick Reference Guide

Analyte	Collection/Storage Container	Preservation (at time of sampling)
Anatoxin-a	Amber glass <i>Avoid exposure to light, as this will degrade the toxin.</i>	Immediately upon collection, freshwater samples should be preserved with 10X Concentrated Sample Diluent to prevent adsorptive loss of toxin. <i>Preservation is necessary for freshwater samples only. Saltwater samples do not require additional reagents for preservation.</i> <i>Avoid exposure to high pH conditions, as this will degrade the toxin.</i>
BMAA	Clear glass Polyethylene terephthalate glycol (PETG) High density polyethylene (HDPE) Polycarbonate (PC) Polypropylene (PP) Polystyrene (PS) <i>Avoid amber glass, as toxin will be lost due to adsorption to container surface.</i>	Freeze <i>Samples should be analyzed immediately or frozen to avoid degradation of toxin.</i>
Cylindrospermopsin	Clear or amber glass Polyethylene terephthalate glycol (PETG) High density polyethylene (HDPE) Polycarbonate (PC) Polypropylene (PP) Polystyrene (PS)	None
Microcystins	Clear or amber glass Polyethylene terephthalate glycol (PETG) <i>Avoid all plastic containers other than PETG, as toxin will be lost due to adsorption to container surface.</i>	None
Saxitoxin	Clear or amber glass Polyethylene terephthalate glycol (PETG) High density polyethylene (HDPE) Polycarbonate (PC) Polypropylene (PP) Polystyrene (PS)	Immediately upon collection, freshwater samples should be preserved with 10X Concentrated Sample Diluent to prevent adsorptive loss of toxin. <i>Preservation is necessary for freshwater samples only. Saltwater samples do not require additional reagents for preservation.</i>

Unless otherwise indicated, samples can be stored refrigerated for up to 5 days. If samples must be held for greater than 5 days, samples should be stored frozen. If samples are to be shipped, they should be shipped overnight, on ice.

R021516

Help Need

Sample from tribal waterbodies experiencing a cyanobacterial bloom: identification of key organisms* **Need your help getting a sample**

1. **Contact me: 407-803-5508; 407-738-0669; brosen@usgs.gov or text 407-738-0669**
2. **Follow standard sampling protocol (see next slide)**
3. **Ship live samples (overnight): Barry Rosen, USGS, 12703 Research Parkway, Orlando, FL 32779**

Thank You!

Cylindrospermopsis raciborskii (CYN)



Cylindrospermopsis sp. (CYN?)



10 μ m

Pseudanabaena contorta (MYC)



Komvophoron sp. (*Pseudanabaena*)



Planktolyngbya limnetica



5 μ m