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Effects of Liming and Nitrogen Fertilization on Emissions of CO₂ and N₂O From a Temperate Forest

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Fluxes of N₂O and CO₂ were measured simultaneously in control, N-fertilized, and limed plots in a 145-year-old beech stand in the Solling area in Germany using an automated chamber method. The N-fertilized plot received annually 140 kg of nitrogen as NH₄SO₄ since 1982; the limed plot was treated with 30 t/ha of dolomitic limestone once in 1982. On all plots, fluxes of N₂O and CO₂ underwent strong diel, daily, and seasonal variations. For the control plot, N₂O and CO₂ effluxes averaged about 1 mg N/m²/d and 2.9 g CO₂/m²/d from October until May and 3 mg N/m²/d and 4.5 g CO₂/m²/d from May until September. Annual fluxes for N₂O and CO₂ were 5.6 kg N/ha/yr and to 3.2 t C/ha/yr, respectively. Nitrogen saturation of the system due to high rates of N deposition for several decades may be responsible for the high N₂O losses. Liming drastically reduced the N₂O emission to 1.5 kg N/ha/yr and increased the CO₂ emission to 4.1 t C/ha/yr, 5 years after the treatment. Changes in N₂O/N₂ ratios are assumed to be the reason for the lower N₂O emission. Fertilization increased the N₂O emission, resulting in 7.8 kg N/ha/yr, whereas the effect on CO₂ emission with 3.8 t C/ha/yr was less pronounced.

1. INTRODUCTION

Temperate forest ecosystems have been characterized until recently as nitrogen deficient. A review of the literature shows that under these conditions the average rates of N₂O release are in the range of 1 kg N₂O-N/ha/yr [Bowden, 1986]. In vast areas of industrialized countries, however, high atmospheric deposition rates of nitric and sulfuric acid have changed the nitrogen and acid status of forest soils [Ulrich, 1983]. In many ecosystems, N saturation has been reached [Beese and Matzner, 1986; Melillo et al., 1989; Aber et al., 1989; Skeffington, 1990], whereby nitrogen deposition exceeds the demand of forests by a factor of 2-8 [Beese and Matzner, 1986]. Under these conditions, increased leaching of nitrate to the groundwater and gaseous N losses, such as nitrous oxide (N₂O) and nitrogen (N₂), may occur. In acid forest soils, nitrous oxide is the main gaseous product of denitrification; however, it is also released as a by-product of the ammonium oxidation (nitrification) [Firestone and Davidson, 1989]. Increased emissions of N₂O and CO₂ from soils to the atmosphere are important because these gases change the chemical and physical properties of the atmosphere. Nitrous oxide enhances the catalytic reduction of ozone [Crutzen, 1974; McElroy et al., 1977], and both contribute to the greenhouse effect by absorbing infrared radiation in the troposphere. Nitrous oxide is of concern because its ability to warm the atmosphere is 180 times greater than that of CO₂ [Lashof and Ahuja, 1990], and it is increasing at 0.2-0.3 %/yr [Rasmussen and Khalil, 1986; Brasseur and Hitchman, 1988].

To prevent further acidification of forest soils and to improve the Ca⁺⁺ and Mg⁺⁺ supply of the trees, increasing areas of forests with acid soils in Germany have been limed

in the last decade. Changes in acidity caused by liming may have an impact on nitrogen cycling and litter decomposition, both of which influence CO₂ and N₂O fluxes. Therefore management-induced changes in forest ecosystems may create side effects, that may influence biosphere-atmospheric interaction. Specifically, increased rates of CO₂ production may result from accelerated decomposition of soil organic matter stored in the forest floor, and increased emission of N₂O may result from higher rates of nitrification and higher NO₃ concentrations in the soil solution.

In this study we attempted to answer the following questions: (1) What are the rates of CO₂ and N₂O emissions from acid forest soils under conditions of high nitrogen input? (2) Do additional NH₄ inputs change CO₂ and N₂O emission rates? (3) Does liming change the CO₂ and N₂O emission rates?

2. STUDY AREA AND TREATMENTS

Measurements of N₂O and CO₂ released from three plots (20 m × 20 m) of a 145-year-old beech stand in the Solling area in Germany were made with three replicates on each plot in the years 1987 and 1988. The limed plot was treated with 30 t/ha of fine ground dolomitic limestone in 1982. The fertilized plot has been fertilized annually with 140 kg of nitrogen in the form of solid ammonium sulfate since 1982. A third plot was taken as control. All plots receive 35 kg N/ha/yr by atmospheric deposition [Matzner, 1989]. The soil is an acidic cambisol derived from loess over weathered triassic sandstone. Some properties of the soil and the humus layer (5-6 cm thick) of the three plots are summarized in Table 1.

3. METHODS

We developed an automatic chamber method for daily monitoring of N₂O and CO₂ fluxes from the soil to the

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TABLE 1. Soil Properties of the Fertilized, Limed, and Control Plots

	Soil, 0-5 cm					Humus Layer					
	pH CaCl ₂ 0.01 M	CEC $\mu\text{mol}_c/\text{g}$	Ca + Mg % of CEC	Organic C %	C/N	O _L		O _r		O _H	
						Organic C %	C/N	Organic C %	C/N	Organic C %	C/N
Control	3.0	129	4.5	5.7	19	51	29	48	22	31	19
Fertilized	3.2	125	3.4	4.1	16	49	26	48	22	31	19
Limed	3.4	128	21.2	5.5	19	49	29	34	21	31	19

atmosphere [Loftfield *et al.*, 1992] to provide reliable rates where high temporal variability of biotic processes occur.

Gas fluxes from the soil into the atmosphere were measured automatically using Plexiglas chambers connected to a gas chromatograph (GC) equipped with an electron capture detector and a personal computer (Figure 1). The chambers consisted of two compartments and were sealed with a movable lid (Figure 2). They were only worked into the litter deep enough to achieve horizontal placement required for the 2-cm water trap to seal the lid. Gas samples were taken from the inner chamber 0, 30, and 60 min after closure. The outer chamber acts as a buffer zone preventing air penetra-

tion from the outside. Each compartment had a volume of 50 L and a surface area of 0.25 m². The personal computer registered opening and closing of the chambers and controlled the extraction of the gas samples, carried out the analysis of samples taken for N₂O and CO₂, the integration of the chromatograms, and the storage of the data. For calibration of the GC a gas standard was analyzed every 3-6 measurements. Each of the nine chambers was closed 5 times each day which resulted in 45 flux calculations in total. Soil temperatures were measured in 5- and 10-cm depth. Air, soil temperatures, and light intensities were recorded as hourly averages. Soil water tension (10-cm depth) were

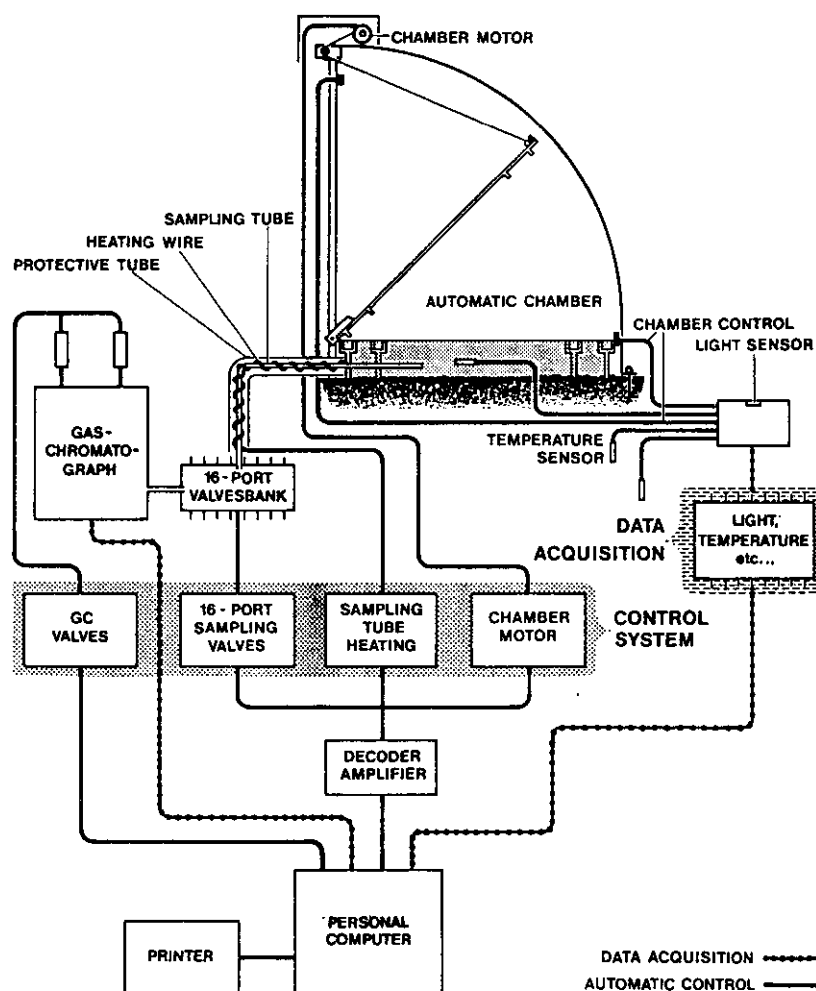


Fig. 1. Schematic diagram of the personal computer, supported control, and data acquisition.

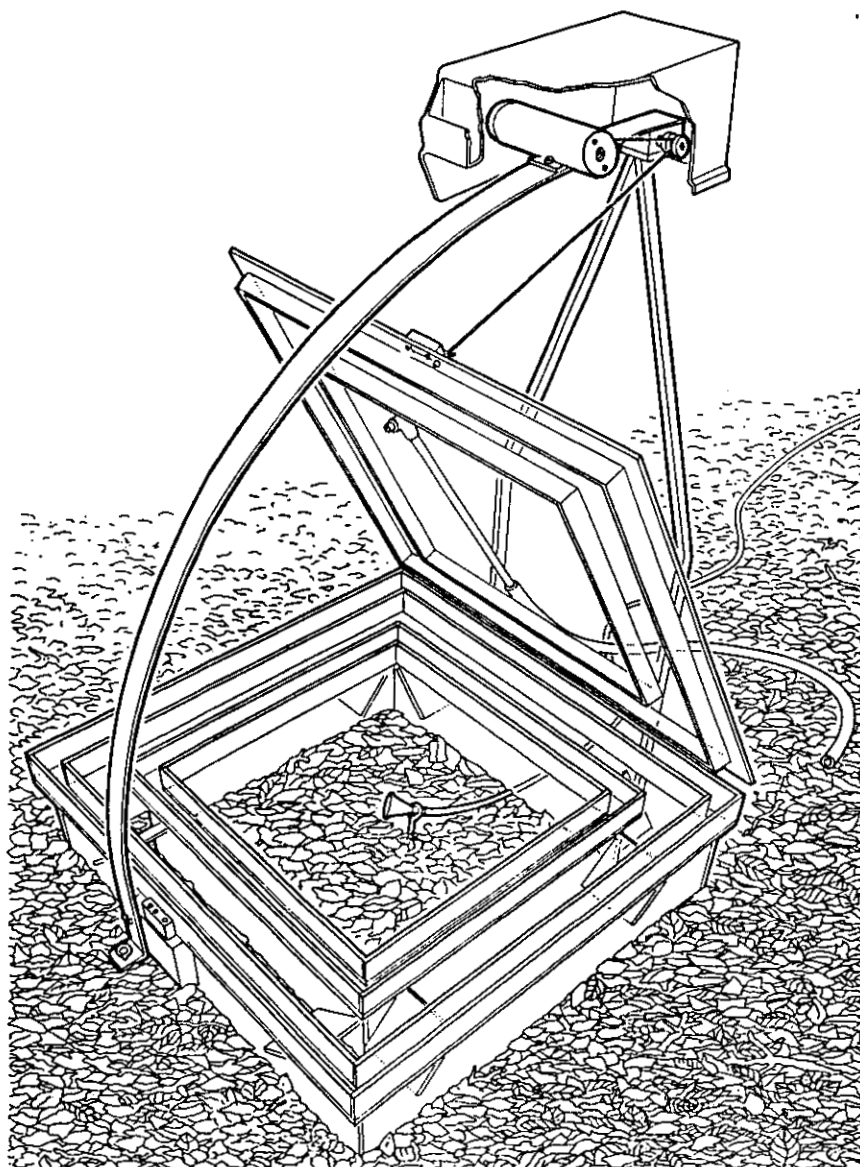


Fig. 2. Lockable double chamber for collecting gaseous losses from soils.

measured once a week and soil water solutions were collected over a month by suction cups in 10-cm depth and were analyzed for nitrate [Meiwes *et al.*, 1984].

4. RESULTS

4.1. Temporal Variations

Nitrous oxide and CO₂ emission showed strong diel and daily variations due to changing environmental conditions (Figure 3). The amplitude of diel fluxes is not constant. As an estimate based on a 20-day plot in June 1988, one can assume that the highest values are 10 to 220% higher for CO₂ and 10 to 470% higher for N₂O than the lowest rate measured each day.

Nitrous oxide emission also underwent a strong seasonal variation. Two main periods of N₂O emission can be distinguished on the control and fertilized plot (Figure 4). On the control plot, low rates (about 1 mg N/m²/d) occurred from

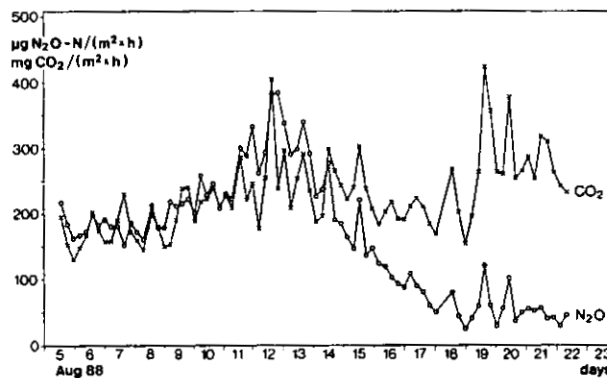


Fig. 3. Diel and daily changes of N₂O and CO₂ emission rates for August 1988 in chamber 6 of the control plot.

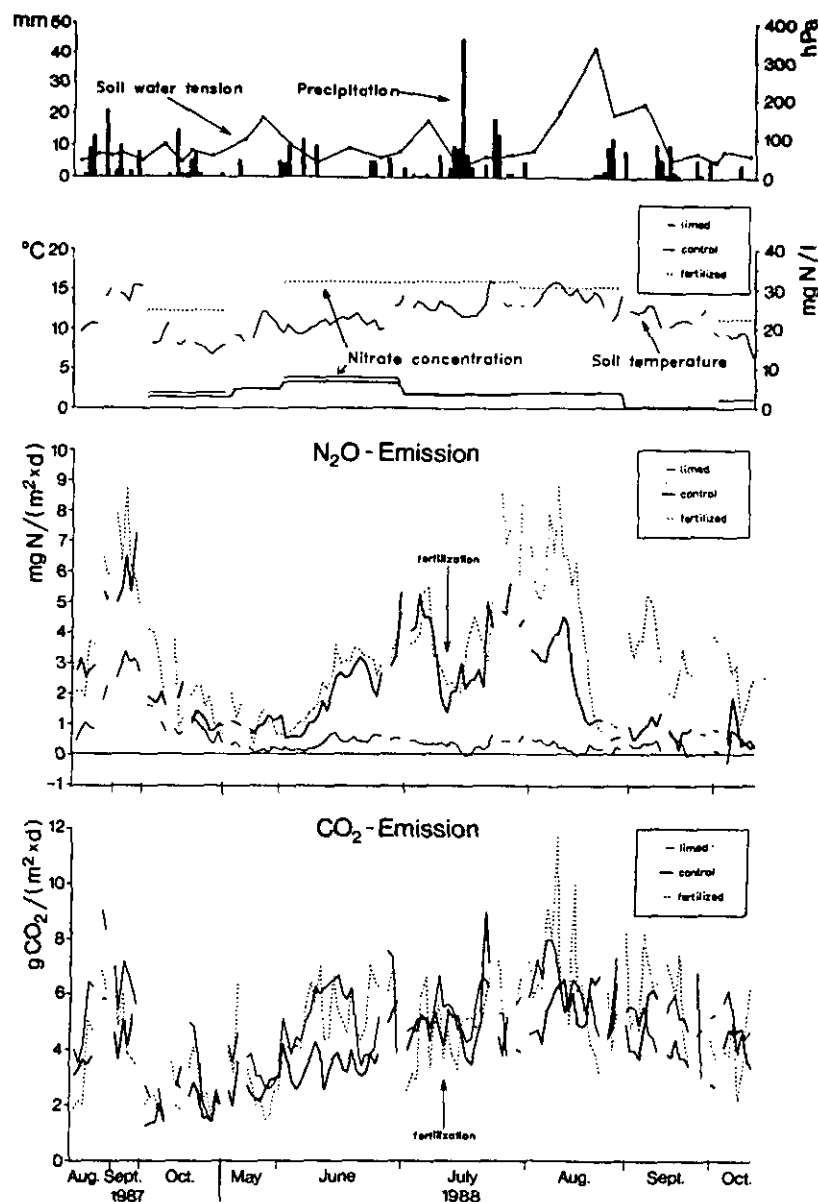


Fig. 4. Daily mean values of CO₂ and N₂O emission of three chambers and monthly nitrate concentration of soil solution (10-cm depth) of three plots, soil water tension (10-cm depth), soil temperature (5-cm depth), and precipitation (interrupted monitoring is indicated by breaks).

September/October to May, and elevated mean rates (approximately 3.5 mg N/m²/d) occurred during the vegetation period. This seasonal fluctuation was not observed on the limed plot. There was only one short period with increased N₂O emission of 3.2 mg N/m²/d at the end of 1987. In 1988 the rates remained below 0.8 mg with an average of 0.5 mg N/m²/d.

Soil respiration rates followed a similar but less-pronounced seasonal pattern, especially on the control plot. Fluctuations due to temperature changing are more pronounced for CO₂, resulting in peaks with a duration of several days. Despite these different patterns of CO₂ and N₂O there are periods with high correlations between both fluxes, for example, in September 1987 and directly after fertilization in 1988.

During the active period of N₂O emission (May to September), precipitation events are often accompanied by lower temperatures and lower emission rates of N₂O on the control and fertilized plot in 1988 (Figure 4). After precipitation periods, often fluxes increased together with soil water tensions and even the temperatures. At the middle of August this period abruptly ended when soil water tension exceeded 200 hPa. The effects of precipitation frequency on the CO₂ emissions were less pronounced.

Fertilization with ammonium sulfate in this nitrifying acid soil [Lang and Beese, 1985] resulted in higher N₂O fluxes than observed on the control plot after a lag time of 14 days (Figure 4). Despite higher nitrate concentrations on the fertilized plot the emission of N₂O prior to the fertilization was only slightly higher than on the control plot. The effect

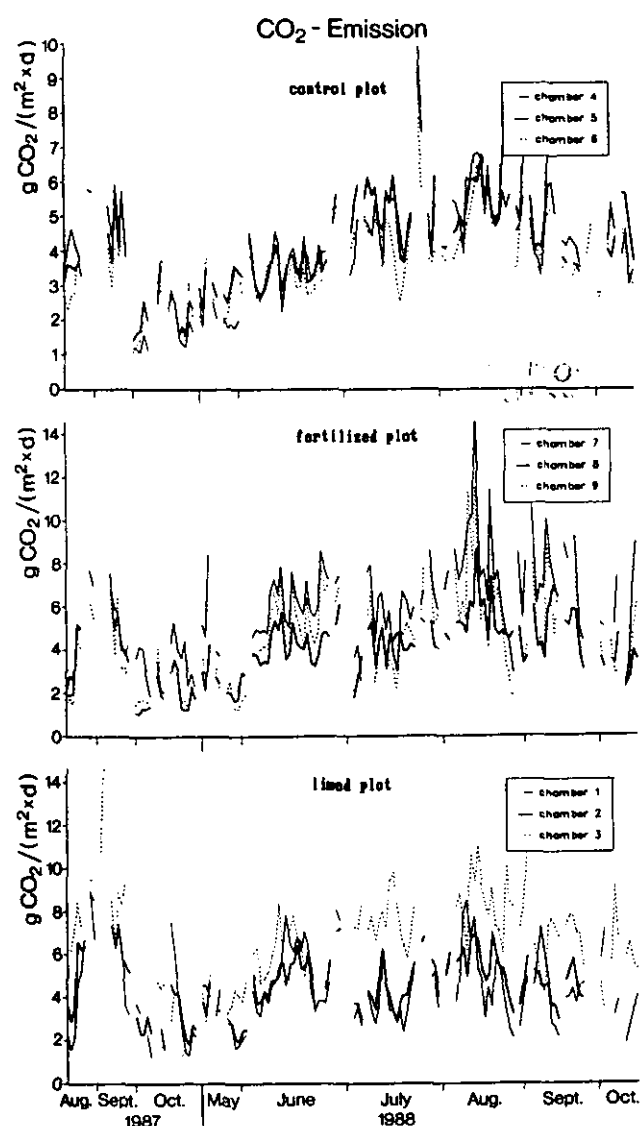


Fig. 5. Daily mean values of CO_2 emission on a control, fertilized, and limed plot of three chambers on each plot.

of fertilization on the CO_2 emission was less pronounced than on N_2O , but the fluctuations were higher than on the other plots.

Temporal variation of CO_2 and N_2O emissions into the three chambers of one plot is very similar (Figures 5 and 6), despite different levels of emission rates among the chambers. The results show that the highest levels of N_2O emission rates of one chamber of a plot were not accompanied by the highest levels of CO_2 .

4.2. Annual Emission Rates of N_2O

Monthly N_2O emissions of the acidic cambisol varied from 0.2 to 1.8 kg N/ha (Table 2). Assuming that the lowest monthly rates occurred during wintertime, emission rates of 4.6, 6.1, and 6.1 kg N/ha/yr from chambers 4, 5, and 6 (Table 3) resulted in a mean rate of 5.6 ± 0.9 kg N/ha/yr.

After fertilization with ammonium sulphate the nitrate concentration in soil solution increased 8 times compared to the control plot (Figure 4). The mean annual emission rate of

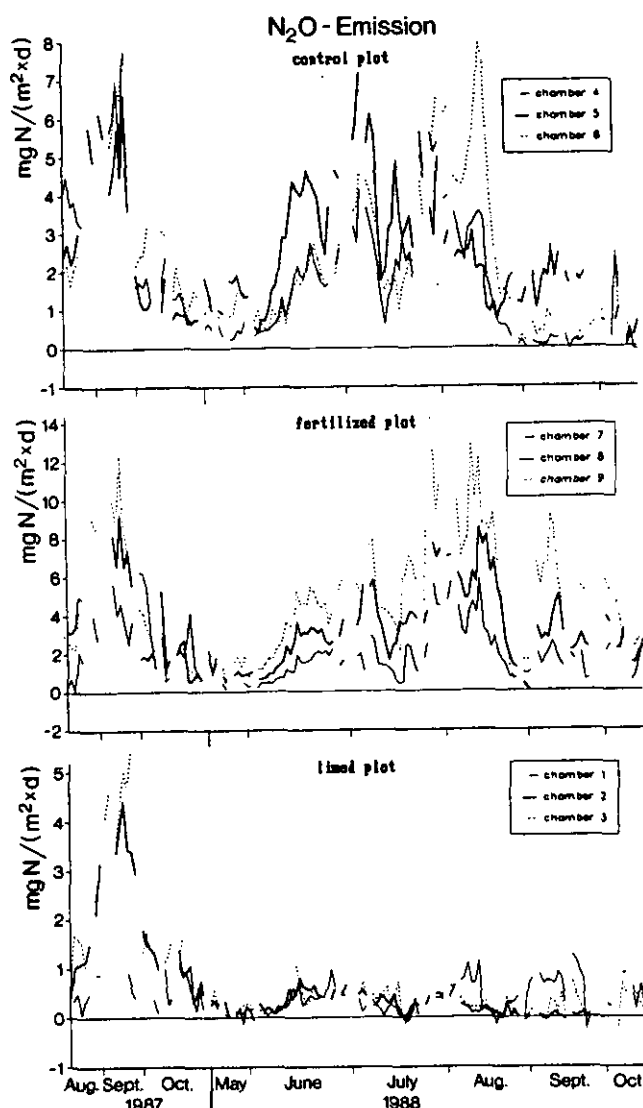


Fig. 6. Daily mean values of N_2O emission on a control, fertilized, and limed plot of three chambers on each plot.

N_2O was increased by 39% to 7.8 ± 2.9 kg N/ha/yr. The variation of the chambers (Table 3, Figure 6) was 2 times higher than that of the control. From the heavy N input of 140 kg N/ha/yr only 1.6% was transformed to N_2O .

Liming drastically reduced the N_2O emissions. Although approximately the same nitrate concentrations were found as in the untreated soil, the N_2O emissions dropped to about 1.5 ± 0.5 kg N/ha/yr, only 26% of the control flux rate. The variations were comparable to the fertilized plot (Table 3).

4.3. Annual Emission Rates of CO_2

The monthly rates of CO_2 from the acidic cambisol varied between 170 and 450 kg C/ha (Table 2). Making the same assumptions as before, an annual emission rate of 3.2 ± 0.3 t C/ha/yr was calculated. The emissions of CO_2 were increased by about 16% on the nitrogen-fertilized plot and about 31% on the limed plot. This resulted in an annual emission of 3.8 ± 1.0 and 4.1 ± 0.9 t C/ha/yr, respectively. The variation between the chambers was significantly reduced compared to N_2O (Table 3).

TABLE 2. Mean Monthly Emission Rates of CO₂ and N₂O on a Control, N Fertilized, and Limed Plot

	Control		Fertilized kg/ha/month		Limed	
	CO ₂ -C	N ₂ O-N	CO ₂ -C	N ₂ O-N	CO ₂ -C	N ₂ O-N
Aug. 1987	332	1.03	325	1.10	510	0.36
Sept. 1987	376	1.78	390	1.96	538	0.88
Oct. 1987	172	0.45	206	0.66	237	0.28
May 1988	221	0.31	254	0.35	288	0.06
June 1988	292	0.57	428	0.70	438	0.11
July 1988	423	1.09	407	1.28	444	0.09
Aug. 1988	454	0.78	535	1.45	524	0.08
Sept. 1988	358	0.24	485	1.04	419	0.08
Oct. 1988	347	0.21	356	0.77	417	0.11

5. DISCUSSION

High temporal variability of trace gas losses from soil to the atmosphere, as indicated by our data, can result in large errors in determination of fluxes with the chamber methods [Mosier, 1989]. Because most of the published emission rates for N₂O and many for CO₂ were determined by single flux determinations once a week. We have used a high temporal resolution to overcome hourly, daily, and seasonal variations. The amplitude of daily fluxes indicates the largest errors using single flux determination per day. As indicated by our data, the differences of the highest daily rates to the lowest were 10 to 470%. To calculate the deviations which may result from single short term flux measurements per week, we have calculated the mean hourly emission rates from 700 measurements (140 days) for each chamber and compared these data with results obtained from 30 measurements (once a week between 0630 and 1130). The results are shown in Table 4. The deviations for the CO₂ emission rates using one determination each week lie between -7% and +11% related to the automatic chambers. The mean deviation is less than 1%. The slight deviation for CO₂ resulted partly from the daytime chosen for the measurements. The deviations for the N₂O emission rates are worse. The deviations lie in the range of +3% to +50% and were on average +21%. This is due to higher rates of N₂O emission between 0630 and 1130. Liming and fertilization resulted in higher deviations compared with the control plot. In general, it was found that regular weekly measurements under the conditions studied resulted in reasonable estimates of the total emissions.

Our measurements exhibit a clear seasonal pattern for N₂O and CO₂ emission rates, upon which are superimposed

daily and hourly variations (Figures 3 and 4). Soil temperature and soil water tension can explain most of the behavior of the emissions. The influence of the controlling factors is basically the same at all subplots (chamber level), indicated by the parallel pattern of emissions (Figures 5 and 6). External factors such as temperature and water probably explain only part of the temporal variation. Additional variations may also be explained by soil factors, such as organic matter or microbial processes.

The annual N₂O flux of 5.6 ± 0.9 kg N/ha released from the acidic cambisol is substantially higher than rates found in other studies. Schmidt *et al.* [1988] only found annual rates between 0.26 and 0.96 kg N/ha on six different deciduous forest stands in Germany. Similar annual rates were found for deciduous forests: 0.47–1.4 kg N/ha [Goodroad *et al.*, 1984] in Wisconsin, 0.15 kg N in New Hampshire [Keller *et al.*, 1983], 0.017 kg for a mixed hardwood stand in Massachusetts [Bowden *et al.*, 1990], and 1 kg N in Massachusetts [Duxbury *et al.*, 1982]. Higher annual rates of 2.4–3.2 kg N/ha were observed in coniferous forest ecosystems in Wisconsin [Goodroad *et al.*, 1984]; however, Bowden *et al.* [1990] showed that coniferous forest ecosystems do not necessarily emit more N₂O than deciduous forests. In a red pine plantation in Massachusetts only 0.01 kg N/ha/yr were emitted. We also detected only low N₂O emission rates in a spruce stand adjacent to the beech stand described in this paper.

The high N₂O emission rates from the Solling site may be caused by the high long term N depositions. Since the beginning of measurements at the Solling site in 1969, yearly nitrogen deposition rates have averaged 35 kg N/ha [Matzner, 1989]. With more or less constant leaching of 5 kg

TABLE 3. Annual Emission Rates of CO₂ and N₂O of Nine Chambers on a Control, N-Fertilized, and Limed Plot

Chamber	Limed			Control			Fertilized		
	1	2	3	4	5	6	7	8	9
t C/ha/yr	3.7	3.6	5.2	3.1	3.5	3.1	4.9	3.2	3.2
cv		21.5%			8.3%			26.5%	
kg N/ha/yr	1.0	1.5	2.0	4.6	6.1	6.1	4.9	7.7	10.7
cv		34.5%			16.2%			37.1%	

Coefficient of variation, cv.

TABLE 4. Mean Hourly Emission Rates of CO₂ and N₂O of Nine Chambers Calculated for the Automatic Chambers Using All Fluxes and for the Case of One Flux Determination Each Week

Chamber	Limed			Control			Fertilized		
	1	2	3	4	5	6	7	8	9
<i>mg CO₂/m²/d</i>									
CO ₂									
automatic chamber*	172	183	266	164	173	149	227	161	172
one flux determination each week†	181	173	279	166	161	141	233	168	190
changes,‡ %	+5.2	-5.5	+4.9	+1.2	-6.9	-5.4	-1.7	+4.3	+10.5
<i>µg N/m²/h</i>									
N ₂ O									
automatic chamber*	16.7	27.3	34.4	84.7	107	93.4	76.4	133	185
one flux determination each week†	22.8	34.6	43.3	87.4	119	103	114	141	219
changes,‡ %	+36.5	+26.7	+25.9	+3.2	+11.2	+10.3	+48.6	+6.0	+18.4

*700 emission rates within 140 days.

†30 emission rates of 30 days within 140 days.

‡One flux determination each week related to automatic chambers.

N as measured separately and losses of 5.7 kg N caused by N₂O emissions, this ecosystem receives 24.3 kg N per year which may be stored in organic matter, trees, or may be lost by N₂.

After fertilization with ammonium the emission rates of N₂O increased after a lag time of 14 days, indicating that nitrification may influence the N₂O losses. Nitrification plays an important role in this acid soil. Heterotrophic microorganisms may be responsible for nitrification, as shown by Lang and Beese [1985]. The higher nitrate concentrations in the soil solution before fertilization seemed to be of minor importance to the N₂O emission. Only 1.6% of the fertilizer nitrogen were transformed to N₂O amounting to a total of 7.8 ± 2.9 kg N/ha/yr. This is in agreement with the results of Bowden *et al.* [1991], who found that after fertilization with 150 kg N/ha/yr as NH₄NO₃ to a mixed hardwood forest and a red pine plantation no more than 0.2% of the added N were transformed to N₂O.

Five years after liming, the N₂O emissions were reduced drastically to annual rates of about 1.5 ± 0.5 kg N/ha/yr. Nitrate concentration in the soil solution was approximately the same as found in the control (Figure 4) but the pH(H₂O) in the humus layer increased from 4.5 to 6.5. These results indicate that the long-term effects of liming result not only in prevention of further acidification of soil by acidic deposition but also in a 74% reduction of N₂O emissions. An explanation for the reduced N₂O release may be found in changed N₂O/N₂ ratios of the gases emitted. Laboratory investigations have shown a shift to higher N₂ proportion after increasing pH [Knowles, 1981].

Estimates of global fluxes of N₂O from temperate deciduous forests covering 4.41 × 10⁶ km² [Steudler *et al.*, 1989; Bowden *et al.*, 1990] amounted to 2.47 Tg N/yr. Schmidt *et al.* [1988] estimated fluxes from temperate forests of 0.7–1.5 Tg N/yr and Bowden *et al.* [1990] a flux of 0.012 Tg N/yr. The results show that areas with higher emissions exist and their contribution to global fluxes must be quantified.

Carbon dioxide release from soils is originally from two different sources, the decomposition of soil organic matter and the respiration of living roots. Both are measured together in the chamber. The annual flux from the control amounts to 3.2 ± 0.3 t C/ha. After fertilization and liming,

the mean rates of CO₂ emission were increased by about 16% to an annual flux of 3.8 ± 1.0 t C/ha and about 31% to 4.1 ± 0.9 t C/ha. These data show that changes in soil chemistry combined with changes in the microbial community may result in accelerated decomposition of organic matter and a reduction of carbon pools of soils. Accelerated CO₂ emission rates on the limed plot were not accompanied by a higher leaching of nitrate (Figure 4).

From these data it is evident that changes in soil chemistry caused by atmospheric deposition (H⁺ and N) or site management drastically may change the emission rates of climate relevant gases. A sensitivity analysis indicates that short time measurements as mostly carried out in the past cannot ever lead to realistic output rates. The data also show that quasi-continuous measurements may improve our knowledge to quantify the influence of the parameters governing the trace gas emission under field conditions. From the data it is obvious that our knowledge of the external and internal soil factors controlling the processes of trace gas emissions is still not good enough for the prognosis of emission rates under changing conditions. This stresses the necessity of using quasi-continuous measurements over longer time periods as a tool for parameter identification and model verification.

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